

Supplementary Material

„Diversity of cilia-based mechanosensory systems and their functions in marine animal behaviour“

Luis Alberto Bezares-Calderón, Jürgen Berger and Gáspár Jékely

Supplementary Table S1. Source data used for filling out the columns in Figure 6.

Taxon	References					
	Directional sensitivity	axoneme	Microvilli (Mv)/ Stereovilli (St)	Lab culture ¹	Transformation ²	Genome ^{3,4}
Nemertea	NK	9+2: apical organ of pilidium larva[1], Proboscis[2]	St: apical organ of pilidium larva[1,3], Ciliary band collar cells[3,4], Proboscis [2]	NK	<i>C. lac.</i> (M)[5] <i>M. ala.</i> (M)[6]	<i>N. gen.</i> [7]
Platyhelminthes	NK	9+2: epidermal receptors[8,9]	No St: epidermal receptors[8] St: epidermal receptors[8], larval epithelium[10]	<i>S. med.</i> , <i>D. jap.</i> [11], <i>M. lig.</i> , <i>S. man.</i> [12]	<i>S. med.</i> , <i>D. jap.</i> (F,E,M)[13], <i>M. lig.</i> (M)[14], <i>S. man.</i> (E,L)[15]	<i>S. med.</i> [16], <i>D. jap.</i> [17], <i>M. med.</i> [18], Tape worms[19], <i>S. man.</i> [20]
Annelida	NK	9+2: CR neurons[21] Modified: statocyst cilia[22]	No St: statocyst cilia[22] St: CR neurons [21]	<i>P. dum.</i> [23–25], <i>C. tel.</i> [26], <i>E. jap.</i> [27] <i>H. rob.</i> [28], <i>Hirudo spp.</i> (see table 2 in [29])	<i>P. dum.</i> [30,31], <i>C. tel.</i> [32], <i>E. jap.</i> [33], <i>H. rob.</i> (all species:M) [34]	<i>C. tel.</i> [35], <i>H. rob.</i> [35], <i>E. fet.</i> [36,37]
Brachiopoda	NK	9+2: collar receptors[38]	Mv: LFC[39], collar receptors[38,40]	NK	NK	<i>L. ana.</i> [41]
Phoronida	NK	9+2: LFC[42–45]	St: LFC[42–47]	NK	NK	<i>P. aus.</i> [7]

Taxon	References					
	Directional sensitivity	axoneme	Microvilli (Mv)/ Stereovilli (St)	Lab culture ¹	Transformation ²	Genome ^{3,4}
Bryozoa	NK	9+2: AFC[48,49]	Mv: AFC[48,49], St: LFC[50]	<i>E. sul.</i> [51]	NK	<i>B. ner.</i> (T)[52]
Gastrotricha	NK	9+2: Statocyst[53], head[54]	St: Statocyst[53], head[54]	<i>L.squ.</i> (C)	NK	<i>L.squ.</i> (T)[55], <i>Meg.</i> (T)[55], <i>D. asp.</i> (T)[55]
Mollusca	NDS.: gastropod statocyst cells [56] DS: cephalopod statocyst cells [57]	9+2: statocyst cilia [58–60], epithelial receptors[61]	No St: statocyst cells[58–60] St: tentacle receptors[61,62]	various (see table 2 in [29])	various (e.g. <i>C. for.</i> (M) [63], <i>L. stag.</i> (M)[64]), bivalves (M/L) [65,66], <i>C. gig.</i> (F)[67]	Several species[68]
Entoprocta	NK	9+2: Apical organ cells [69]	St: Apical organ cells[69]	NK	NK	<i>B. elo.</i> (T)[70]
Chaetognatha	NK	9+2: ciliary fence [71,72]	St: ciliary fence [71,73]	Various (e.g. <i>P. got.</i> , <i>S. his.</i>) [74–76]	<i>P. got.</i> (M)[77]	various[78, 79]
Rotifera	NK	9+2: touch bristles[80], anterior sensory endings[80], antennae[80] 9+0: palpar organ [80]	No St: touch bristles[80], antennal receptors[81]	<i>B. pli./man.</i> , <i>A. sie.</i>	<i>B. pli./man.</i> (L/E)[82,83]	Various species [84–87]
Micrognathozoa	NK	9+0: sensory bristles [88]	NK	NK	NK	<i>L. mae.</i> (T) [89]
Gnathostomulida	NK	9+2: ciliary cirri [90,91] Spiral ciliary	St: ciliary cirri [90,91] (referred by [94])	NK	NK	Various (T) [89]

Taxon	References					
	Directional sensitivity	axoneme	Microvilli (Mv)/ Stereovilli (St)	Lab culture ¹	Transformation ²	Genome ^{3,4}
		organ[92,93]				
Arthropoda	DS: bristle receptors[95]	9+0: chordotonal receptors[96] Mod: campaniform sensilla[96]	NoSt: Type I receptor cells	Several species (marine <i>P. haw.</i> [97])	Several species (F,M, E)	Several species
Onychophora	NK	Mod: antenna bipolar cell[98], upside-down cell[98]	St: upside-down cell[98]	NK	NK	<i>E. row.</i> (T)[99, 100]
Tardigrada	NK	Mod: cirri ciliary cells[101], anterior-lateral sensory field, sub-oral sensory region [102]	NoSt: cirri ciliary cells[101]	NK	<i>H. exe</i> (M)[103]	<i>H. exe</i> [104]. <i>R. var.</i> [105]
Nematoda	NK	9+0: metaneme proprioceptor[106] Modified: labial and setal mechanoreceptors[107–109], Mechanosensillum [110],	NoSt: labial and setal mechanoreceptors[107,108] St(like): metaneme proprioceptor [106]	Not widespread (only one report found[111])	Various species (M,B,F,S)[112–114]	Several species[115]
Nematomorpha	NK	NK	NK	<i>P. var.</i> [116]	NK	<i>Gordius</i> sp.(T)[117]
Loricifera	NK	NK	St: N-flosculi[118,119]	NK	NK	<i>A. ele.</i> (T) [89]

Taxon	References					
	Direct- ional sensiti- vity	axoneme	Microvilli (Mv)/ Stereovilli (St)	Lab cult- ure ¹	Trans- formation ²	Genome ^{3,4}
Kino- rhyncha	NK	9+0: Sensory spot Type I & II cells[120] Mod: scalid type I[121–123] 9+2: sensory spot receptor [124]	St: Sensory spot type I cell [120,122] NoSt: scalid type I[121,122], sensory spot type II cell[120]	NK	NK	<i>Pycnophyes</i> sp. (T)[117]
Priapulida	NK	9+2: epithelial receptors [125,126] 9+0: epithelial receptors (distally)[125], type 2 receptors [127]	St: epithelial receptors[125,126], scalid receptors [127,128]	NK	NK	<i>H. spi.</i> (T) [129], <i>T. cor.</i> (T)[89]
Vertebrata	DS: Hair cells	9+2: hair cells	St: hair cells	Several species	Several species	Several species
Tunicata	DS(tentative): graded length of stereovilli [130]	9+2: coronal sensory cell [130] Mod: Langherans receptors [131]	No St: coronal sensory cell[130], Langherans receptors [131] St: coronal sensory cell [130]	Various species (e.g. <i>Ciona</i> [132], <i>O.dio.</i> [133,134], <i>D. geg.</i> [135])	Various species (M,E)	Various species [136](e.g. <i>C. int.</i> [137], <i>O. dio.</i> [138], <i>S. tho</i> [139])
Cephalo- chordata	NK	9+2: Type 1 cell[140] Mod: Type 2 cell[140,141]	No St.: oral spine cell[142] St: Type I cell[141]	Various species [143–145]	<i>Branch. spp.</i> (M)[145,146]	<i>Branch. spp.</i> [147,148]
Hemi- chordata	NK	9+2: tentacle cilia[149]	Mv: tentacle cilia[149]	NK	<i>S.kov.</i> (M) [150,151]	<i>S. kov./P. fla.</i> [152]

Taxon	References					
	Directional sensitivity	axoneme	Microvilli (Mv)/ Stereovilli (St)	Lab culture ¹	Transformation ²	Genome ^{3,4}
Echino-dermata	NK	9+2: ciliary band cells[153] Ciliated cells pedicellariae[154,155]	Mv: ciliary band cells[153] Podial sensory receptors[156] Ciliated cells pedicellariae[154,155] NoSt: sea cucumber statocyst cells [157]	Various species (C)[158,159]	Various species (M)[160]	Various species [161]
Xenacoelomorpha	DS (tentative): Structural asymmetry of stereovilli [162,163]	9+2: Epidermal sensory cells[162,163]	No St: Type 1 epidermal sensory cells [162] St: Type 2 epidermal sensory cells [162,163]	<i>H. mia.</i> [164] <i>I. pul.</i> , <i>C. mac.</i> [165] <i>S. ros.</i> [166]	<i>H. mia.</i> (M, S), <i>I. pul.</i> (S)[167], <i>N. fus.</i> [168]	Various species (T)[169]
Cnidaria	DS: hair bundles[170] DS(tentative): C, F & T cells [171,172] NDS: Cnidocytes[173]	9+2: C, F & T cells [171,172], statocyst cells hair bundle cell[174] Mod: nematocyte[175], Concentric hair cells[176]	NoSt: Type A statocyst sensory cells[177] St: nematocyte C, F & T cells [171,172] hair bundle cell[174] proprioceptor [178]	Various species (e.g. <i>N. vec.</i> [179], <i>C. hem.</i> [180])	Various species (S, M, E)[181–185],	Various species[186]
Placozoa	NK	NK	NK	<i>T. adh.</i> [187]	<i>T. adh.</i> (S)[188]	<i>T. adh.</i> [189]
Ctenophora	DS: balancer cilia[190]	9+2: Balancer cilia[190] Tastborsten[191]	No St: Balancer cilia[190] St(like): actin peg[191]	Various species[192]	<i>M. lei.</i> (M)	<i>P. bac.</i> [193], <i>M. lei.</i> [194]
Porifera	NK	9+0: osculum biciliated cells [195]	(Mv): apopylar cells [196]	various species[197]	<i>T. wil.</i> (F) [198], <i>E. mue.</i> (F)[198], <i>S.</i>	<i>A. que.</i> [200], <i>S. car.</i> , <i>X. tes.</i> [201]

Taxon	References					
	Directional sensitivity	axoneme	Microvilli (Mv)/ Stereovilli (St)	Lab culture ¹	Transformation ²	Genome ^{3,4}
				(e.g. <i>S. dom.</i> , <i>E. mue.</i>)	<i>dom.</i> (L) [199]	
Choanoflagellates*	NK	9+2: flagellum	NK	<i>S. rose.</i> [202]	<i>S. rose.</i> (E)[202]	<i>M. bre.</i> [203], <i>S. ros.</i> [204], various species (T) [205]

NOTES:

¹ Marine species where the complete life cycle can be reproduced in the lab are included. Non-marine aquatic species are underscored. If no culturable aquatic species exist for a particular taxon, species that can be easily obtained from commercial or other suppliers throughout the year are also included (C). Seasonal species are marked (S).

² Representative reports in any species of a given phylum are included. Transformation methods considered: microinjection (M), biobalistics (B), electroporation (E), lipofection (L), feeding (F), or soaking (S). Reports in non-aquatic species are included as they may spur development in aquatic/marine species.

³ Only species with assembled and published genomes are included. If no genome sequence is available for a given taxon, species with sequenced transcriptome (T) are included.

⁴ List of sequenced animal genomes available in Wikipedia:
https://en.wikipedia.org/wiki/List_of_sequenced_animal_genomes

*No evidence to our knowledge of mechanosensitivity of the choanoflagellate cilium.

Species abbreviations

A. ele.: *Armorialoricus elegans*; **A. que.:** *Amphimedon queenslandica*; **A. sie.:** *Asplanchna sieboldi*; **B. elo.:** *Barentsia elongata*; **B. ner.:** *Bugula neritina*; **B. pli./man.:** *Brachionus plicatilis/manjivas*; **Branch. spp.:** *Branchiostoma spp.*; **B. ste.:** *Berghia stephanieae*; **C.for.:** *Crepidula fornicata*; **C.gig.:** *Crassostrea gigas*; **C.int.:** *Ciona intestinalis*; **C.lac.:** *Cerebratulus lacteus*; **C.mac.:** *Convolutiloba macropyga*; **D.asp.:** *Diuronotus aspetos*; **D.geg.:** *Dolioletta gegenbauri*; **D.jap.:** *Dugesia japonica*; **E.jap.:** *Enchytraeus japonensis*; **E.fet.:** *Eisenia fetida*; **E.mue.:** *Ephydatia muelleri*; **E.row.:** *Euperipatoides rowelli*; **F.sul.:** *Fredericella sultana*; **H.exe.(duj.:** *Hypsibius exemplaris (dujardini)*; **Hirudo spp.:** *Hirudo spp.*; **H.mia.:** *Hofstenia miamia*; **H.rob.:** *Helobdella robusta*; **H.spi.:** *Halicryptus spinulosus*; **H.spp.:** *Hirudo spp.*; **I.pul.:** *Isodiametra pulchra*; **L.ana.:** *Lingula anatina*; **L.mae.:** *Limnognathia maerski*; **L.squ.:** *Lepidodermella squamata*; **L.sta.:** *Lymnaea stagnalis*; **Meg.:** *Megadasyus sp.*; **M.ala.:** *Macaulaura alaskensis*; **M.lei.:** *Mnemiopsis leidyi*; **M.lig.:** *Macrostromum lignano*; **N.fus.:** *Neochildia fusca*; **N.gen.:** *Notospermus geniculatus*; **N.vec.:** *Nematostella vectensis*; **O.dio.:** *Oikopleura dioica*; **P.aus.:** *Phoronis australis*; **P.bac.:** *Pleurobrachia bachei*; **P fla.:** *Ptychodera flava*; **P.got.:** *Paraspadella gotoi*; **P.haw.:** *Parhyale hawaiiensis*; **P.var.:** *Paragordius varius*; **R.var.:** *Ramazzottius varieornatus*; **S.car.:** *Stylissa carteri*; **S.dom.:** *Suberites domuncula*; **S.kow.:** *Saccoglossus kowalevskii*; **S.man.:** *Schistosoma mansonii*; **S.his.:** *Sagitta hispida*; **S.med.:** *Schmidtea mediterranea*; **S.pur.:** ; **S.ros.:** *Symsagittifera roscoffensis*; **S.rose.:** *Salpingoeca rosetta*; **S.tho.:** *Salpa thompsoni*; **T.cor.:** *Tubiluchus cf. corallicola*; **T.wil.:** *Tethya wilhelma*; **X.tes.:** *Xestospingia testudinaria*

Other abbreviations: **AFC:** abfrontal cilia; **DS:** directionally sensitive; **LFC:** laterofrontal cilia, **NDS:** not directionally sensitive, **NK:** not to our knowledge

References

- Chernyshev, A. V., Astakhova, A. A., Dautov, S. S. & Yushin, V. V. 2013 The morphology of the apical organ and adjacent epithelium of pilidium prorecurvatum, a pelagic larva of an unknown heteronemertean (Nemertea). *Russ. J. Mar. Biol.* **39**, 116–124. (doi:10.1134/S106307401302003X)
- Montalvo, S., Junoy, J., Roldán, C. & García-Corrales, P. 1996 Ultrastructural study of sensory cells of the proboscoidal glandular epithelium of *Riseriellus occultus* (Nemertea, Heteronemertea). *J. Morphol.* **229**, 83–96. (doi:10.1002/(SICI)1097-4687(199607)229:1<83::AID-JMOR5>3.0.CO;2-Q)
- Lacalli, T. C. & West, J. E. 1985 The nervous system of a pilidium larva: evidence from electron microscope reconstructions. *Can. J. Zool.* **63**, 1909–1916. (doi:10.1139/z85-284)
- von Dassow, G., Emler, R. B. & Maslakova, S. A. 2013 How the pilidium larva feeds. *Front. Zool.* **10**, 47. (doi:10.1186/1742-9994-10-47)
- Henry, J. J. & Martindale, M. Q. 1998 Conservation of the spiralian developmental program: cell lineage of the nemertean, *Cerebratulus lacteus*. *Dev. Biol.* **201**, 253–269. (doi:10.1006/dbio.1998.8966)
- von Dassow, G. & Maslakova, S. A. 2017 The trochoblasts in the pilidium larva break an ancient spiralian constraint to enable continuous larval growth and maximally indirect development. *Evodevo* **8**, 19. (doi:10.1186/s13227-017-0079-5)
- Luo, Y.-J., Kanda, M., Koyanagi, R., Hisata, K., Akiyama, T., Sakamoto, H., Sakamoto, T. & Satoh, N. 2018 Nemertean and phoronid genomes reveal lophotrochozoan evolution and the origin of bilaterian heads. *Nat. Ecol. Evol.* **2**, 141–151. (doi:10.1038/s41559-017-0389-y)
- Ehlers, U. & Ehlers, B. 1977 Monociliary receptors in interstitial Proseriata and Neorhabdocoela (Turbellaria Neoophora). *Zoomorphologie* **86**, 197–222. (doi:10.1007/BF00993666)
- De León, R. P. & Volonteri, O. 2003 First description of collar receptors in Temnocephalidae (Platyhelminthes). *Acta Zoologica* **84**, 155–160. (doi:10.1046/j.1463-6395.2003.00141.x)
- Rohde, K., Watson, N. A. & Lanfranchi, A. 1991 Secondary sensory receptor in a platyhelminth. *Naturwissenschaften* **78**, 463–465. (doi:10.1007/BF01134385)
- Merryman, M. S., Alvarado, A. S. & Jenkin, J. C. 2018 Culturing Planarians in the Laboratory. *Methods Mol. Biol.* **1774**, 241–258. (doi:10.1007/978-1-4939-7802-1_5)
- Mann, V. H., Morales, M. E., Rinaldi, G. & Brindley, P. J. 2010 Culture for genetic manipulation of developmental stages of *Schistosoma mansoni*. *Parasitology* **137**, 451–462. (doi:10.1017/S0031182009991211)
- Rouhana, L., Weiss, J. A., Forsthoefel, D. J., Lee, H., King, R. S., Inoue, T., Shibata, N., Agata, K. & Newmark, P. A. 2013 RNA interference by feeding in vitro-synthesized double-stranded RNA to planarians: methodology and dynamics. *Dev. Dyn.* **242**, 718–730. (doi:10.1002/dvdy.23950)
- Wudarski, J. et al. 2017 Efficient transgenesis and annotated genome sequence of the regenerative flatworm model *Macrostomum lignano*. *Nat. Commun.* **8**, 2120. (doi:10.1038/s41467-017-02214-8)
- Correnti, J. M., Jung, E., Freitas, T. C. & Pearce, E. J. 2007 Transfection of *Schistosoma mansoni* by electroporation and the description of a new promoter sequence for transgene expression. *Int. J. Parasitol.* **37**, 1107–1115. (doi:10.1016/j.ijpara.2007.02.011)
- Grohme, M. A. et al. 2018 The genome of *Schmidtea mediterranea* and the evolution of core cellular mechanisms. *Nature* **554**, 56–61. (doi:10.1038/nature25473)
- An, Y. et al. 2018 Draft genome of *Dugesia japonica* provides insights into conserved regulatory elements of the brain restriction gene *nou-darake* in planarians. *Zoological Lett.* **4**, 24. (doi:10.1186/s40851-018-0102-2)
- Wasik, K. et al. 2015 Genome and transcriptome of the regeneration-competent flatworm, *Macrostomum lignano*. *Proc. Natl. Acad. Sci. USA* **112**, 12462–12467. (doi:10.1073/pnas.1516718112)
- Tsai, I. J. et al. 2013 The genomes of four tapeworm species reveal adaptations to parasitism. *Nature* **496**, 57–63. (doi:10.1038/nature12031)
- Berriman, M. et al. 2009 The genome of the blood fluke *Schistosoma mansoni*. *Nature* **460**, 352–358. (doi:10.1038/nature08160)
- Bezares-Calderón, L. A., Berger, J., Jasek,

- S., Verasztó, C., Mendes, S., Gühmann, M., Almeida, R., Shahidi, R. & Jékely, G. 2018 Neural circuitry of a polycystin-mediated hydrodynamic startle response for predator avoidance. *Elife* **7**. (doi:10.7554/eLife.36262)
22. Locke, J. M. 2000 Ultrastructure of the statocyst of the marine enchytraeid *Grania americana* (Annelida: Clitellata). *Invertebr. Biol.* **119**, 83–93. (doi:10.1111/j.1744-7410.2000.tb00176.x)
23. Hauenschild, C. & Fischer, A. 1969 *Platynereis dumerilii*: mikroskopische Anatomie, Fortpflanzung, Entwicklung. Stuttgart: G. Fischer, 1969.
24. Kuehn, E., Stockinger, A. W., Girard, J., Raible, F. & Özpolat, B. D. 2019 A scalable culturing system for the marine annelid *Platynereis dumerilii*. *BioRxiv* (doi:10.1101/714238)
25. Veedin-Rajan, V. B., Fischer, R. M., Raible, F. & Tessmar-Raible, K. 2013 Conditional and specific cell ablation in the marine annelid *Platynereis dumerilii*. *PLoS One* **8**, e75811. (doi:10.1371/journal.pone.0075811)
26. Elaine Seaver Lab 2014 The Ultimate Capitella teleta Culturing Protocol.
27. Kawamoto, S., Yoshida-Noro, C. & Tochinai, S. 2005 Bipolar head regeneration induced by artificial amputation in *Enchytraeus japonensis* (Annelida, Oligochaeta). *J. Exp. Zool. A. Comp. Exp. Biol.* **303**, 615–627. (doi:10.1002/jez.a.205)
28. Weisblat, D. A. & Kuo, D.-H. 2009 Handling of *Helobdella* (leech) embryos. *Cold Spring Harb. Protoc.* **2009**, pdb.prot5189. (doi:10.1101/pdb.prot5189)
29. Henry, J. Q. 2014 Spiralian model systems. *Int. J. Dev. Biol.* **58**, 389–401. (doi:10.1387/ijdb.140127jh)
30. Zantke, J., Bannister, S., Rajan, V. B. V., Raible, F. & Tessmar-Raible, K. 2014 Genetic and genomic tools for the marine annelid *Platynereis dumerilii*. *Genetics* **197**, 19–31. (doi:10.1534/genetics.112.148254)
31. Ackermann, C. F. 2003 Markierung der Zelllinien im Embryo von *Platynereis* - Deutsche Digitale Bibliothek.
32. Neal, S., de Jong, D. M. & Seaver, E. C. 2019 CRISPR/CAS9 mutagenesis of a single r-opsin gene blocks phototaxis in a marine larva. *Proc. Biol. Sci.* **286**, 20182491. (doi:10.1098/rspb.2018.2491)
33. Takeo, M., Yoshida-Noro, C. & Tochinai, S. 2010 Functional analysis of grimp, a novel gene required for mesodermal cell proliferation at an initial stage of regeneration in *Enchytraeus japonensis* (Enchytraeidae, Oligochaeta). *Int. J. Dev. Biol.* **54**, 151–160. (doi:10.1387/ijdb.082790mt)
34. Zhang, S. O. & Weisblat, D. A. 2005 Applications of mRNA injections for analyzing cell lineage and asymmetric cell divisions during segmentation in the leech *Helobdella robusta*. *Development* **132**, 2103–2113. (doi:10.1242/dev.01802)
35. Simakov, O. et al. 2013 Insights into bilaterian evolution from three spiralian genomes. *Nature* **493**, 526–531. (doi:10.1038/nature11696)
36. Paul, S., Arumugaperumal, A., Rathy, R., Ponesakki, V., Arunachalam, P. & Sivasubramaniam, S. 2018 Data on genome annotation and analysis of earthworm *Eisenia fetida*. *Data Brief* **20**, 525–534. (doi:10.1016/j.dib.2018.08.067)
37. Springett, J. A. 1964 A method for culturing enchytraeidae. *Oikos* **15**, 175. (doi:10.2307/3564754)
38. Lüter, C. 1996 The median tentacle of the larva of *lingula anatina* (brachiopoda) from queensland, australia. *Aust J Zool* **44**, 355. (doi:10.1071/ZO9960355)
39. Gilmour, T. H. J. 1981 Food-collecting and waste-rejecting mechanisms in *Glottidia pyramidata* and the persistence of lingulacean inarticulate brachiopods in the fossil record. *Can. J. Zool.* **59**, 1539–1547. (doi:10.1139/z81-209)
40. Lüter, C. 2015 Brachiopoda. In *Structure and evolution of invertebrate nervous systems*, pp. 341–350. Oxford University Press. (doi:10.1093/acprof:oso/9780199682201.003.0027)
41. Luo, Y.-J. et al. 2015 The Lingula genome provides insights into brachiopod evolution and the origin of phosphate biomineralization. *Nat. Commun.* **6**, 8301. (doi:10.1038/ncomms9301)
42. Pardos, F., Roldán, C., Benito, J. & Emig, C. C. 1991 Fine Structure of the Tentacles of *Phoronis australis* Haswell (Phoronida, Lophophorata). *Acta Zoologica* **72**, 81–90. (doi:10.1111/j.1463-6395.1991.tb00320.x)
43. Gilmour, T. H. J. 1978 Ciliation and function of the food-collecting and waste-rejecting organs of lophophorates. *Can. J. Zool.* **56**, 2142–2155. (doi:10.1139/z78-290)
44. Fernández, I., Pardos, F., Benito, J. & Roldán, C. 1996 Ultrastructural observations on the phoronid nervous system. *J Morphol* **230**, 265–281. (doi:10.1002/(SICI)1097-4687(199612)230:3<265::AID-JMOR2>3.0.CO;2-D)
45. Pardos, F., Roldán, C., Benito, J., Aguirre, A. & Fernández, I. 1993 Ultrastructure of the lophophoral tentacles in the genus *Phoronis* (Phoronida, Lophophorata). *Can. J. Zool.* **71**, 1861–1868. (doi:10.1139/z93-265)
46. Hay-Schmidt, A. 1989 The nervous system of the actinotroch larva of *Phoronis muelleri* (Phoronida). *Zoomorphology* **108**, 333–351.

- (doi:10.1007/BF00312274)
47. Lacalli, T. C. 1990 Structure and organization of the nervous system in the actinotroch larva of phoronis vancouverensis. *Philos. Trans. R. Soc. Lond. B, Biol. Sci.* **327**, 655–685. (doi:10.1098/rstb.1990.0104)
 48. Shunatova, N. N. & Nielsen, C. 2005 Putative sensory structures in marine bryozoans. *Invertebr. Biol.* **121**, 262–270. (doi:10.1111/j.1744-7410.2002.tb00066.x)
 49. Tamberg, Y. & Shunatova, N. 2017 Tentacle structure in freshwater bryozoans. *J Morphol* **278**, 718–733. (doi:10.1002/jmor.20666)
 50. Nielsen, C. & Riisgård, H. U. 1998 Tentacle structure and filter-feeding in *Crisia eburnea* and other cyclostomatous bryozoans, with a review of upstream-collecting mechanisms. *Mar. Ecol. Prog. Ser.* **168**, 163–186. (doi:10.3354/meps168163)
 51. Grabner, D. S. & El-Matbouli, M. 2008 Transmission of Tetracapsuloides bryosalmonae (Myxozoa: Malacosporea) to *Fredericella sultana* (Bryozoa: Phylactolaemata) by various fish species. *Dis. Aquat. Organ.* **79**, 133–139. (doi:10.3354/dao01894)
 52. Wong, Y. H., Ryu, T., Seridi, L., Ghosheh, Y., Bougouffa, S., Qian, P.-Y. & Ravasi, T. 2014 Transcriptome analysis elucidates key developmental components of bryozoan lophophore development. *Sci. Rep.* **4**, 6534. (doi:10.1038/srep06534)
 53. Marotta, R., Todaro, M. A. & Ferraguti, M. 2008 The unique gravireceptor organs of *Pleurodasys helgolandicus* (Gastrotricha: Macrodasyida). *Zoomorphology* **127**, 111–119. (doi:10.1007/s00435-008-0056-6)
 54. Hochberg, R. 2001 A special form of sensory cilia in *Lepidodermella squamata* (Gastrotricha: Chaetonotida). *Ophelia* **55**, 137–139. (doi:10.1080/00785236.2001.10409480)
 55. Fromm, B., Tosar, J. P., Aguilera, F., Friedländer, M. R., Bachmann, L. & Hejnol, A. 2019 Evolutionary Implications of the microRNA- and piRNA Complement of *Lepidodermella squamata* (Gastrotricha). *Noncoding RNA* **5**. (doi:10.3390/ncrna5010019)
 56. Wolff, H. G. 1972 Multi-directional sensitivity of statocyst receptor cells of the opisthobranch gastropod *Aplysia limacina*. *Mar. Behav. Physiol.* **1**, 361–373. (doi:10.1080/10236247209386910)
 57. Budelmann, B. U. & Williamson, R. 1994 Directional sensitivity of hair cell afferents in the Octopus statocyst. *J. Exp. Biol.* **187**, 245–259.
 58. Barber, V. C. 1966 The fine structure of the statocyst of *Octopus vulgaris*. *Z. Zellforsch.* **70**, 91–107. (doi:10.1007/BF00345067)
 59. Barber, V. C. & Dilly, P. N. 1969 Some aspects of the fine structure of the statocysts of the molluscs *Pecten* and *Pterotrachea*. *Z. Zellforsch.* **94**, 462–478. (doi:10.1007/BF00936053)
 60. Geuze, J. J. 1967 Observations on the function and the structure of the statocysts of *lymnaea stagnalis* (L.). *Neth. J. Zool.* **18**, 155–204. (doi:10.1163/002829668X00090)
 61. Crisp, M. 1981 Epithelial sensory structures of trochids. *J. Mar. Biol. Ass.* **61**, 95. (doi:10.1017/S0025315400045938)
 62. Moir, A. J. 1977 Ultrastructural studies on the ciliated receptors of the long tentacles of the giant scallop, *Placopecten magellanicus* (gmelin). *Cell Tissue Res.* **184**, 367–380. (doi:10.1007/bf00219897)
 63. Perry, K. J. & Henry, J. Q. 2015 CRISPR/Cas9-mediated genome modification in the mollusc, *Crepidula fornicata*. *Genesis* **53**, 237–244. (doi:10.1002/dvg.22843)
 64. Abe, M., Shimizu, M. & Kuroda, R. 2009 Expression of exogenous fluorescent proteins in early freshwater pond snail embryos. *Dev. Genes Evol.* **219**, 167–173. (doi:10.1007/s00427-009-0278-8)
 65. Cadoret, J. P., Gendreau, S., Delecheneau, J. M., Rousseau, C. & Mialhe, E. 1997 Microinjection of bivalve eggs: application in genetics. *Mol Marine Biol Biotechnol* **6**, 72–77.
 66. Guerra, R. & Esponda, P. 2006 Transfection of eggs in the bivalve mollusc *Chamelea gallina* (Bivalvia, Veneridae). *J Submicrosc Cytol Pathol* **38**, 5–10.
 67. Feng, D., Li, Q. & Yu, H. 2019 RNA Interference by Ingested dsRNA-Expressing Bacteria to Study Shell Biosynthesis and Pigmentation in *Crassostrea gigas*. *Mar. Biotechnol.* **21**, 526–536. (doi:10.1007/s10126-019-09900-2)
 68. Takeuchi, T. 2017 Molluscan genomics: implications for biology and aquaculture. *Curr. Mol. Biol. Rep.* **3**, 297–305. (doi:10.1007/s40610-017-0077-3)
 69. Sensenbaugh, T. 1987 Ultrastructural Observations on the Larva of *Loxosoma pectinaria* Franzén (Entoprocta, Loxosomatidae). *Acta Zoologica* **68**, 135–145. (doi:10.1111/j.1463-6395.1987.tb00884.x)
 70. Hausdorf, B., Helmkamp, M., Meyer, A., Witek, A., Herlyn, H., Bruchhaus, I., Hankeln, T., Struck, T. H. & Lieb, B. 2007 Spiralian phylogenomics supports the resurrection of Bryozoa comprising Ectoprocta and Entoprocta. *Mol. Biol. Evol.* **24**, 2723–2729. (doi:10.1093/molbev/msm214)
 71. Müller, C. H. G., Rieger, V., Perez, Y. & Harzsch, S. 2014 Immunohistochemical and ultrastructural studies on ciliary sense

- organs of arrow worms (Chaetognatha). *Zoomorphology* **133**, 167–189. (doi:10.1007/s00435-013-0211-6)
72. Horridge, G. A. & Boulton, P. S. 1967 Prey Detection by Chaetognatha via a Vibration Sense. *Proceedings of the Royal Society B: Biological Sciences* **168**, 413–419. (doi:10.1098/rspb.1967.0072)
73. Bone, Q. & Pulsford, A. 1978 The arrangement of ciliated sensory cells in Spadella (Chaetognatha). *J. Mar. Biol. Ass.* **58**, 565. (doi:10.1017/S0025315400041229)
74. Harzsch, S., Müller, C. H. G. & Perez, Y. 2015 Chaetognatha. In *Evolutionary developmental biology of invertebrates 1* (ed A. Wanninger), pp. 215–240. Vienna: Springer Vienna. (doi:10.1007/978-3-7091-1862-7_10)
75. Reeve, M. R. 1970 Complete cycle of development of a pelagic chaetognath in culture. *Nature* **227**, 381. (doi:10.1038/227381a0)
76. Goto, T. & Yoshida, M. 1997 Growth and reproduction of the benthic arrowworm *Paraspadella gotoi* (Chaetognatha) in laboratory culture. *Invertebr. Reprod. Dev.* **32**, 201–207. (doi:10.1080/07924259.1997.9672625)
77. Shimotori, T. & Goto, T. 1999 Establishment of Axial Properties in the Arrow Worm Embryo, *Paraspadella gotoi* (Chaetognatha): Developmental Fate of the First Two Blastomeres. *Zool Sci* **16**, 459–469. (doi:10.2108/zsj.16.459)
78. Marlétaz, F., Peijnenburg, K. T. C. A., Goto, T., Satoh, N. & Rokhsar, D. S. 2019 A New Spiralian Phylogeny Places the Enigmatic Arrow Worms among Gnathiferans. *Curr. Biol.* **29**, 312–318.e3. (doi:10.1016/j.cub.2018.11.042)
79. Marlétaz, F., Gilles, A., Caubit, X., Perez, Y., Dossat, C., Samain, S., Gyapay, G., Wincker, P. & Le Parco, Y. 2008 Chaetognath transcriptome reveals ancestral and unique features among bilaterians. *Genome Biol.* **9**, R94. (doi:10.1186/gb-2008-9-6-r94)
80. Clément, P., Wurdak, E. & Amsellem, J. 1983 Behavior and ultrastructure of sensory organs in rotifers. In *Biology of Rotifers* (eds B. Pejler R. Starkweather & T. Nogrady), pp. 89–129. Dordrecht: Springer Netherlands. (doi:10.1007/978-94-009-7287-2_13)
81. Clément, P. & Wurdak, E. 1991 Rotifera. In *Microscopic Anatomy Of Invertebrates, Aschelminthes* (eds F. W. Harrison & E. E. Ruppert), pp. 219–296. New York: Wiley-Liss Inc.
82. Shearer, T. L. & Snell, T. W. 2007 Transfection of siRNA into Brachionus plicatilis (Rotifera). *Hydrobiologia* **593**, 141–150. (doi:10.1007/s10750-007-9067-4)
83. Snell, T. W., Shearer, T. L. & Smith, H. A. 2011 Exposure to dsRNA elicits RNA interference in Brachionus manjavacas (Rotifera). *Mar. Biotechnol.* **13**, 264–274. (doi:10.1007/s10126-010-9295-x)
84. Kim, H.-S. et al. 2018 The genome of the freshwater monogonont rotifer Brachionus calyciflorus. *Mol. Ecol. Resour.* **18**, 646–655. (doi:10.1111/1755-0998.12768)
85. Nowell, R. W. et al. 2018 Comparative genomics of bdelloid rotifers: Insights from desiccating and nondesiccating species. *PLoS Biol.* **16**, e2004830. (doi:10.1371/journal.pbio.2004830)
86. Han, J., Park, J. C., Choi, B.-S., Kim, M.-S., Kim, H.-S., Hagiwara, A., Park, H. G., Lee, B.-Y. & Lee, J.-S. 2019 The genome of the marine monogonont rotifer Brachionus plicatilis: Genome-wide expression profiles of 28 cytochrome P450 genes in response to chlorpyrifos and 2-ethyl-phenanthrene. *Aquat. Toxicol.* **214**, 105230. (doi:10.1016/j.aquatox.2019.105230)
87. Flot, J.-F. et al. 2013 Genomic evidence for ameiotic evolution in the bdelloid rotifer Adineta vaga. *Nature* **500**, 453–457. (doi:10.1038/nature12326)
88. Kristensen, R. M. & Funch, P. 2000 Micrognathozoa: a new class with complicated jaws like those of Rotifera and Gnathostomulida. *J. Morphol.* **246**, 1–49. (doi:10.1002/1097-4687(200010)246:1<1::AID-JMOR1>3.0.CO;2-D)
89. Laumer, C. E. et al. 2015 Spiralian phylogeny informs the evolution of microscopic lineages. *Curr. Biol.* **25**, 2000–2006. (doi:10.1016/j.cub.2015.06.068)
90. Lemmert, V. 1991 Gnathostomulida. In *Microscopic Anatomy Of Invertebrates, Aschelminthes* (eds F. W. Harrison & E. E. Ruppert), pp. 19–39. Wiley-liss.
91. Lammert, V. 1986 Ciliäre Rezeptoren der Gnathostomulida. *Verh. Dtsch. Zool. Ges.*, 283–284.
92. Kristensen, R. M. & Nørrevang, A. 1977 On the Fine Structure of Rastrognathia macrostoma gen. et sp.n. Placed in Rastrognathiidae fam.n. (Gnathostomulida). *Zool Scr* **6**, 27–41. (doi:10.1111/j.1463-6409.1977.tb00757.x)
93. Lammert, V. 1984 The fine structure of spiral ciliary receptors in Gnathostomulida. *Zoomorphology* **104**, 360–364. (doi:10.1007/BF00312186)
94. Schmidt-Rhaesa, A. 2007 Sensory organs. In *The evolution of organ systems*, pp. 118–147. Oxford University Press. (doi:10.1093/acprof:oso/9780198566687.003.0007)
95. Walker, R. G., Willingham, A. T. & Zuker, C.

- S. 2000 A *Drosophila* mechanosensory transduction channel. *Science* **287**, 2229–2234.
96. Keil, T. A. 1997 Functional morphology of insect mechanoreceptors. *Microsc. Res. Tech.* **39**, 506–531. (doi:10.1002/(SICI)1097-0029(19971215)39:6<506::AID-JEMT5>3.0.CO;2-B)
97. Sun, D. A. & Patel, N. H. 2019 The amphipod crustacean *Parhyale hawaiiensis*: An emerging comparative model of arthropod development, evolution, and regeneration. *Wiley Interdiscip. Rev. Dev. Biol.* **8**, e355. (doi:10.1002/wdev.355)
98. Storch, V. & Ruhberg, H. 1977 Fine structure of the sensilla of *Peripatopsis moseleyi* (Onychophora). *Cell Tissue Res.* **177**, 539–553. (doi:10.1007/bf00220613)
99. Treffkorn, S. & Mayer, G. 2013 Expression of the decapentaplegic ortholog in embryos of the onychophoran *Euperipatoides rowelli*. *Gene Expr. Patterns* **13**, 384–394. (doi:10.1016/j.gep.2013.07.004)
100. Treffkorn, S., Kahnke, L., Hering, L. & Mayer, G. 2018 Expression of NK cluster genes in the onychophoran *Euperipatoides rowelli*: implications for the evolution of NK family genes in nephrozoans. *Evodevo* **9**, 17. (doi:10.1186/s13227-018-0105-2)
101. Kristensen, R. M. 1981 Sense organs of two marine arthrotardigrades (heterotardigrada, tardigrada). *Acta Zoologica* **62**, 27–41. (doi:10.1111/j.1463-6395.1981.tb00614.x)
102. Walz, B. 1978 Electron microscopic investigation of cephalic sense organs of the tardigrade *Macrobiotus hufelandi* C.A.S. Schultze. *Zoomorphologie* **89**, 1–19. (doi:10.1007/BF00993778)
103. Tenlen, J. R. 2018 Microinjection of dsRNA in Tardigrades. *Cold Spring Harb. Protoc.* **2018**. (doi:10.1101/pdb.prot102368)
104. Yoshida, Y. et al. 2017 Comparative genomics of the tardigrades *Hypsibius dujardini* and *Ramazzottius varieornatus*. *PLoS Biol.* **15**, e2002266. (doi:10.1371/journal.pbio.2002266)
105. Hashimoto, T. et al. 2016 Extremotolerant tardigrade genome and improved radiotolerance of human cultured cells by tardigrade-unique protein. *Nat. Commun.* **7**, 12808. (doi:10.1038/ncomms12808)
106. Hope, W. D. & Gardiner, S. L. 1982 Fine structure of a proprioceptor in the body wall of the marine nematode *Deontostoma californicum* Steiner and Albin, 1933 (Enoplida: Leptosomatidae). *Cell Tissue Res.* **225**, 1–10. (doi:10.1007/BF00216213)
107. Wright, K. A. 1977 Labial sense organs of the nematode, *Heterakis gallinarum*. *J. Parasitol.* **63**, 528–539.
108. Croll, N. A. & Smith, J. M. 1974 Nematode setae as mechanoreceptors. *Nematol.* **20**, 291–296. (doi:10.1163/187529274X00339)
109. Doroquez, D. B., Berciu, C., Anderson, J. R., Sengupta, P. & Nicastro, D. 2014 A high-resolution morphological and ultrastructural map of anterior sensory cilia and glia in *Caenorhabditis elegans*. *Elife* **3**, e01948. (doi:10.7554/eLife.01948)
110. Wright, K. A. 1992 Peripheral sensilla of some lower invertebrates: the Platyhelminthes and Nematoda. *Microsc. Res. Tech.* **22**, 285–297. (doi:10.1002/jemt.1070220306)
111. Moens, T. & Vincx, M. 1998 On the cultivation of free-living marine and estuarine nematodes. *Helgoländer Meeresunters.* **52**, 115–139. (doi:10.1007/BF02908742)
112. Tabara, H., Grishok, A. & Mello, C. C. 1998 RNAi in *C. elegans*: soaking in the genome sequence. *Science* **282**, 430–431. (doi:10.1126/science.282.5388.430)
113. Timmons, L. & Fire, A. 1998 Specific interference by ingested dsRNA. *Nature* **395**, 854. (doi:10.1038/27579)
114. Fire, A., Xu, S., Montgomery, M. K., Kostas, S. A., Driver, S. E. & Mello, C. C. 1998 Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. *Nature* **391**, 806–811. (doi:10.1038/35888)
115. Sommer, R. J. & Streit, A. 2011 Comparative genetics and genomics of nematodes: genome structure, development, and lifestyle. *Annu. Rev. Genet.* **45**, 1–20. (doi:10.1146/annurev-genet-110410-132417)
116. Hanelt, B. & Janovy Jr, J. 2004 Untying a Gordian knot: the domestication and laboratory maintenance of a Gordian worm, *Paragordius varius* (Nematomorpha: Gordiida). *J. Nat. Hist.* **38**, 939–950. (doi:10.1080/0022293021000058718)
117. Smythe, A. B., Holovachov, O. & Kocot, K. M. 2019 Improved phylogenomic sampling of free-living nematodes enhances resolution of higher-level nematode phylogeny. *BMC Evol. Biol.* **19**, 121. (doi:10.1186/s12862-019-1444-x)
118. Schmidt-Rhaesa, A., editor 2012 *Nematomorpha, priapulida, kinorhyncha, loricifera*. Berlin, Boston: DE GRUYTER. (doi:10.1515/9783110272536)
119. Kristensen, R. M. 1991 Loricifera. In *Aschelminthes. Microscopic anatomy of invertebrates* (eds F. W. Harrison & E. E. Ruppert), pp. 351–375. New York: Wiley-Liss.
120. Nebelsick, M. 1992 Sensory Spots of *Echinoderes capitatus* (Zelinka, 1928) (Kinorhyncha, Cyclorhagida). *Acta Zoologica* **73**, 185–195. (doi:10.1111/j.1463-6395.1992.tb01186.x)
121. Moritz, K. & Storch, V. 1972 Über den

- ultrastrukturellen bau der Skaliden von Trachydemus giganteus (Kinorhyncha). *Mar. Biol.* **16**, 81–89. (doi:10.1007/BF00347852)
122. Brown, R. 1986 Developmental and taxonomic studies of Sydney Harbour kinorhyncha.
123. Brown, R. 1989 Morphology and ultrastructure of the sensory appendages of a kinorhynch introvert. *Zool Scr* **18**, 471–482. (doi:10.1111/j.1463-6409.1989.tb00141.x)
124. Kristensen, R. M. & Higgins, P. 1991 Kinorhyncha. In *Aschelminthes. Microscopic anatomy of invertebrates* (eds F. W. Harrison & E. E. Ruppert), pp. 377–404. New York.: Wiley-Liss.
125. Moritz, K. & Storch, V. 1971 Elektronenmikroskopische Untersuchung eines Mechanorezeptors von Evertabraten (Priapuliden, Oligochaeten). *Z. Zellforsch.* **117**, 226–234. (doi:10.1007/BF00330739)
126. Higgins, R. P., Storch, V. & Shirley, T. C. 1993 Scanning and Transmission Electron Microscopical Observations on the Larvae of *Priapulus caudatus* (Priapulida). *Acta Zoologica* **74**, 301–319. (doi:10.1111/j.1463-6395.1993.tb01245.x)
127. Storch, V., Higgins, R. P., Anderson, P., Svavarsson, J. & Svavarsson, J. 1995 Scanning and Transmission Electron Microscopic Analysis of the Introvert of *Priapulopsis australis* and *Priapulopsis bicaudatus* (Priapulida). *Invertebr. Biol.* **114**, 64. (doi:10.2307/3226954)
128. Storch, V., Higgins, R. P., Malakhov, V. V. & Adrianov, A. V. 1994 Microscopic anatomy and ultrastructure of the introvert of *Priapulus caudatus* and *P. tuberculatospinosus* (Priapulida). *J Morphol* **220**, 281–293. (doi:10.1002/jmor.1052200307)
129. Borner, J., Rehm, P., Schill, R. O., Ebersberger, I. & Burmester, T. 2014 A transcriptome approach to ecdysozoan phylogeny. *Mol. Phylogenet. Evol.* **80**, 79–87. (doi:10.1016/j.ympev.2014.08.001)
130. Burighel, P., Caicci, F. & Manni, L. 2011 Hair cells in non-vertebrate models: lower chordates and molluscs. *Hear. Res.* **273**, 14–24. (doi:10.1016/j.heares.2010.03.087)
131. Bone, Q. & Ryan, K. P. 1979 The Langerhans receptor of *Oikopleura* (Tunicata: Larvacea). *J. Mar. Biol. Ass.* **59**, 69. (doi:10.1017/S002531540004618X)
132. Joly, J.-S., Kano, S., Matsuoka, T., Auger, H., Hirayama, K., Satoh, N., Awazu, S., Legendre, L. & Sasakura, Y. 2007 Culture of *Ciona intestinalis* in closed systems. *Dev. Dyn.* **236**, 1832–1840. (doi:10.1002/dvdy.21124)
133. Nishida, H. 2008 Development of the appendicularian *Oikopleura dioica*: culture, genome, and cell lineages. *Dev. Growth Differ.* **50 Suppl 1**, S239–56. (doi:10.1111/j.1440-169X.2008.01035.x)
134. Martí-Solans, J. et al. 2015 *Oikopleura dioica* culturing made easy: a low-cost facility for an emerging animal model in EvoDevo. *Genesis* **53**, 183–193. (doi:10.1002/dvg.22800)
135. Deibel, D. 1982 Laboratory determined mortality, fecundity and growth rates of *Thalia democratica* Forskal and *Dolioletta gegenbauri* Uljanin (Tunicata, Thaliacea). *J. Plankton Res.* **4**, 143–153. (doi:10.1093/plankt/4.1.143)
136. Berná, L. & Alvarez-Valin, F. 2014 Evolutionary genomics of fast evolving tunicates. *Genome Biol. Evol.* **6**, 1724–1738. (doi:10.1093/gbe/evu122)
137. Dehal, P. et al. 2002 The draft genome of *Ciona intestinalis*: insights into chordate and vertebrate origins. *Science* **298**, 2157–2167. (doi:10.1126/science.1080049)
138. Denoeud, F. et al. 2010 Plasticity of animal genome architecture unmasked by rapid evolution of a pelagic tunicate. *Science* **330**, 1381–1385. (doi:10.1126/science.1194167)
139. Jue, N. K., Batta-Lona, P. G., Trusiak, S., Obergfell, C., Bucklin, A., O'Neill, M. J. & O'Neill, R. J. 2016 Rapid Evolutionary Rates and Unique Genomic Signatures Discovered in the First Reference Genome for the Southern Ocean Salp, *Salpa thompsoni* (Urochordata, Thaliacea). *Genome Biol. Evol.* **8**, 3171–3186. (doi:10.1093/gbe/evw215)
140. Bone, Q. & Best, A. C. G. 1978 Ciliated sensory cells in amphioxus (branchiostoma). *J. Mar. Biol. Ass.* **58**, 479. (doi:10.1017/S0025315400028137)
141. Lacalli, T. C. & Hou, S. 1999 A reexamination of the epithelial sensory cells of amphioxus (Branchiostoma). *Acta Zoologica* **80**, 125–134. (doi:10.1046/j.1463-6395.1999.80220005.x)
142. Lacalli and, T. C., Gilmour, T. H. J. & Kelly, S. J. 1999 The oral nerve plexus in amphioxus larvae: function, cell types and phylogenetic significance. *Proc. R. Soc. Lond. B* **266**, 1461–1470. (doi:10.1098/rspb.1999.0801)
143. Carvalho, J. E., Lahaye, F. & Schubert, M. 2017 Keeping amphioxus in the laboratory: an update on available husbandry methods. *Int. J. Dev. Biol.* **61**, 773–783. (doi:10.1387/ijdb.170192ms)
144. Holland, L. Z. & Yu, J.-K. 2004 Cephalochordate (amphioxus) embryos: procurement, culture, and basic methods. In *Development of sea urchins, ascidians, and other invertebrate deuterostomes: experimental approaches*, pp. 195–215. Elsevier. (doi:10.1016/S0091-679X(04)74009-1)

145. Le Petillon, Y., Bertrand, S. & Escrivà, H. 2020 Spawning Induction and Embryo Micromanipulation Protocols in the Amphioxus *Branchiostoma lanceolatum*. *Methods Mol. Biol.* **2047**, 347–359. (doi:10.1007/978-1-4939-9732-9_19)
146. Hirsinger, E., Carvalho, J. E., Chevalier, C., Lutfalla, G., Nicolas, J.-F., Peyri  ras, N. & Schubert, M. 2015 Expression of fluorescent proteins in *Branchiostoma lanceolatum* by mRNA injection into unfertilized oocytes. *J. Vis. Exp.*, 52042. (doi:10.3791/52042)
147. Marl  taz, F. et al. 2018 Amphioxus functional genomics and the origins of vertebrate gene regulation. *Nature* **564**, 64–70. (doi:10.1038/s41586-018-0734-6)
148. Putnam, N. H. et al. 2008 The amphioxus genome and the evolution of the chordate karyotype. *Nature* **453**, 1064–1071. (doi:10.1038/nature06967)
149. Dilly, P. N. 1972 The structures of the tentacles of *Rhabdopleura compacta* (Hemichordata) with special reference to neurociliary control. *Z. Zellforsch. Mikrosk. Anat.* **129**, 20–39.
150. Lowe, C. J. et al. 2006 Dorsoventral patterning in hemichordates: insights into early chordate evolution. *PLoS Biol.* **4**, e291. (doi:10.1371/journal.pbio.0040291)
151. Lowe, C. J., Tagawa, K., Humphreys, T., Kirschner, M. & Gerhart, J. 2004 Hemichordate embryos: procurement, culture, and basic methods. In *Development of sea urchins, ascidians, and other invertebrate deuterostomes: experimental approaches*, pp. 171–194. Elsevier.(doi:10.1016/S0091-679X(04)74008-X)
152. Simakov, O. et al. 2015 Hemichordate genomes and deuterostome origins. *Nature* **527**, 459–465. (doi:10.1038/nature16150)
153. Nerrevang, A. & Wingstrand, K. G. 1970 On the Occurrence and Structure of Choanocyte-like Cells in Some Echinoderms. *Acta Zoologica* **51**, 249–270. (doi:10.1111/j.1463-6395.1970.tb00436.x)
154. Peters, B. H. & Campbell, A. C. 1987 Morphology of the nervous and muscular systems in the heads of pedicellariae from the sea urchin *Echinus esculentus* L. *J Morphol* **193**, 35–51. (doi:10.1002/jmor.1051930105)
155. Cobb, J. L. S. 1968 The fine structure of the pedicellariae of *Echinus esculentus* (L.). *Journal of the Royal Microscopical Society* **88**, 223–233. (doi:10.1111/j.1365-2818.1968.tb00610.x)
156. Burke, R. D. 1980 Podial sensory receptors and the induction of metamorphosis in echinoids. *J. Exp. Mar. Bio. Ecol.* **47**, 223–234. (doi:10.1016/0022-0981(80)90040-4)
157. Ehlers, U. 1997 Ultrastructure of the Statocysts in the Apodous Sea Cucumber *Leptosynapta inhaerens* (Holothuroidea, Echinodermata). *Acta Zoologica* **78**, 61–68. (doi:10.1111/j.1463-6395.1997.tb01127.x)
158. Foltz, K. R., Adams, N. L. & Runft, L. L. 2004 Echinoderm eggs and embryos: procurement and culture. In *Development of sea urchins, ascidians, and other invertebrate deuterostomes: experimental approaches*, pp. 39–74. Elsevier.(doi:10.1016/S0091-679X(04)74003-0)
159. Hodin, J. et al. 2019 Culturing echinoderm larvae through metamorphosis. *Methods Cell Biol.* **150**, 125–169. (doi:10.1016/bs.mcb.2018.11.004)
160. Kiehart, D. P. 1982 Microinjection of echinoderm eggs: apparatus and procedures. *Methods Cell Biol.* **25 Pt B**, 13–31. (doi:10.1016/s0091-679x(08)61418-1)
161. Kudtarkar, P. & Cameron, R. A. 2017 Echinobase: an expanding resource for echinoderm genomic information. *Database (Oxford)* **2017**. (doi:10.1093/database/bax074)
162. Bedini, C., Ferrero, E. & Lanfranchi, A. 1973 The ultrastructure of ciliary sensory cells in two *Turbellaria Acoela*. *Tissue Cell* **5**, 359–372. (doi:10.1016/s0040-8166(73)80030-8)
163. Pfisterm  ller, R. & Tyler, S. 2002 Correlation of fluorescence and electron microscopy of F-actin-containing sensory cells in the epidermis of *Convoluta pulchra* (Platyhelminthes: Acoela). *Acta Zoologica* **83**, 15–24. (doi:10.1046/j.1463-6395.2002.00095.x)
164. Srivastava, M., Mazza-Curll, K. L., van Wolfswinkel, J. C. & Reddien, P. W. 2014 Whole-body acoel regeneration is controlled by Wnt and Bmp-Admp signaling. *Curr. Biol.* **24**, 1107–1113. (doi:10.1016/j.cub.2014.03.042)
165. Shannon, T. & Achatz, J. G. 2007 *Convolutriloba macropyga* sp. nov., an uncommonly fecund acoel (Acoelomorpha) discovered in tropical aquaria. *Zootaxa* **1525**, 1–17. (doi:10.11646/zootaxa.1525.1.1)
166. Bailly, X. et al. 2014 The chimerical and multifaceted marine acoel *Symsagittifera roscoffensis*: from photosymbiosis to brain regeneration. *Front. Microbiol.* **5**, 498. (doi:10.3389/fmicb.2014.00498)
167. De Mulder, K. et al. 2009 Characterization of the stem cell system of the acoel *Isodiametra pulchra*. *BMC Dev. Biol.* **9**, 69. (doi:10.1186/1471-213X-9-69)
168. Henry, J. Q., Martindale, M. Q. & Boyer, B. C. 2000 The unique developmental program of the acoel flatworm, *Neochildia fusca*. *Dev. Biol.* **220**, 285–295. (doi:10.1006/dbio.2000.9628)

169. Cannon, J. T., Vellutini, B. C., Smith, J., Ronquist, F., Jondelius, U. & Hejnol, A. 2016 Xenacoelomorpha is the sister group to Nephrozoa. *Nature* **530**, 89–93. (doi:10.1038/nature16520)
170. Mire, P. & Watson, G. M. 1997 Mechanotransduction of hair bundles arising from multicellular complexes in anemones. *Hear. Res.* **113**, 224–234. (doi:10.1016/S0378-5955(97)00145-7)
171. Arkett, S. A., Mackie, G. O. & Meech, R. W. 1988 Hair Cell Mechanoreception in the Jellyfish *Aequorea victoria*. *J. Exp. Biol.*
172. Singla, C. L. 1983 Fine structure of the sensory receptors of *Aequorea victoria* (Hydromedusae: Trachylina). *Cell Tissue Res.* **231**, 415–425. (doi:10.1007/BF00222191)
173. Brinkmann, M., Oliver, D. & Thurm, U. 1996 Mechanoelectric transduction in nematocytes of a hydrozoan (Cnidaria). *J. Comp. Physiol. A* **178**. (doi:10.1007/BF00189597)
174. Peteya, D. J. 1975 The ciliary-cone sensory cell of anemones and cerianthids. *Tissue Cell* **7**, 243–252. (doi:10.1016/0040-8166(75)90003-8)
175. Slautterback, D. B. 1967 The cnidoblast-musculoepithelial cell complex in the tentacles of hydra. *Z. Zellforsch.* **79**, 296–318. (doi:10.1007/BF00369292)
176. Holtmann, M. & Thurm, U. 2001 Variations of concentric hair cells in a Cnidarian sensory epithelium (*Coryne tubulosa*). *J. Comp. Neurol.* **432**, 550–563. (doi:10.1002/cne.1119)
177. Singla, C. L. 1975 Statocysts of hydromedusae. *Cell Tissue Res.* **158**. (doi:10.1007/BF00223835)
178. Peteya, D. J. 1973 A possible proprioceptor in *Ceriantheopsis americanus* (Cnidaria, ceriantharia). *Z. Zellforsch.* **144**, 1–10. (doi:10.1007/BF00306682)
179. Stefanik, D. J., Friedman, L. E. & Finnerty, J. R. 2013 Collecting, rearing, spawning and inducing regeneration of the starlet sea anemone, *Nematostella vectensis*. *Nat. Protoc.* **8**, 916–923. (doi:10.1038/nprot.2013.044)
180. Houlston, E., Momose, T. & Manuel, M. 2010 *Clytia hemisphaerica*: a jellyfish cousin joins the laboratory. *Trends Genet.* **26**, 159–167. (doi:10.1016/j.tig.2010.01.008)
181. Jakob, W. & Schierwater, B. 2007 Changing hydrozoan bauplans by silencing Hox-like genes. *PLoS One* **2**, e694. (doi:10.1371/journal.pone.0000694)
182. Jones, V. A. S., Bucher, M., Hambleton, E. A. & Guse, A. 2018 Microinjection to deliver protein, mRNA, and DNA into zygotes of the cnidarian endosymbiosis model *Aiptasia* sp. *Sci. Rep.* **8**, 16437. (doi:10.1038/s41598-018-34773-1)
183. Momose, T., De Cian, A., Shiba, K., Inaba, K., Giovannangeli, C. & Concordet, J.-P. 2018 High doses of CRISPR/Cas9 ribonucleoprotein efficiently induce gene knockout with low mosaicism in the hydrozoan *Clytia hemisphaerica* through microhomology-mediated deletion. *Sci. Rep.* **8**, 11734. (doi:10.1038/s41598-018-30188-0)
184. Karabulut, A., He, S., Chen, C.-Y., McKinney, S. A. & Gibson, M. C. 2019 Electroporation of short hairpin RNAs for rapid and efficient gene knockdown in the starlet sea anemone, *Nematostella vectensis*. *Dev. Biol.* **448**, 7–15. (doi:10.1016/j.ydbio.2019.01.005)
185. Layden, M. J., Röttinger, E., Wolenski, F. S., Gilmore, T. D. & Martindale, M. Q. 2013 Microinjection of mRNA or morpholinos for reverse genetic analysis in the starlet sea anemone, *Nematostella vectensis*. *Nat. Protoc.* **8**, 924–934. (doi:10.1038/nprot.2013.009)
186. Technau, U. & Schwaiger, M. 2015 Recent advances in genomics and transcriptomics of cnidarians. *Mar. Genomics* **24 Pt 2**, 131–138. (doi:10.1016/j.margen.2015.09.007)
187. Schierwater, B. 2005 My favorite animal, *Trichoplax adhaerens*. *Bioessays* **27**, 1294–1302. (doi:10.1002/bies.20320)
188. Jakob, W., Sagasser, S., Dellaporta, S., Holland, P., Kuhn, K. & Schierwater, B. 2004 The *Trox-2* Hox/ParaHox gene of *Trichoplax* (Placozoa) marks an epithelial boundary. *Dev. Genes Evol.* **214**, 170–175. (doi:10.1007/s00427-004-0390-8)
189. Srivastava, M. et al. 2008 The *Trichoplax* genome and the nature of placozoans. *Nature* **454**, 955–960. (doi:10.1038/nature07191)
190. Tamm, S. L. 1982 Ctenophora. In *Electrical conduction and behaviour in "simple" invertebrates* (ed G. A. B. Shelton), pp. 266–358. Oxford: Clarendon Press.
191. Tamm, S. & Tamm, S. 1991 Actin pegs and ultrastructure of presumed sensory receptors of *Beroë* (Ctenophora). *Cell Tissue Res.* **264**, 151–159. (doi:10.1007/BF00305733)
192. Patry, W. L., Bubel, M. K., Hansen, C. & Knowles, T. 2019 Diffusion tubes: a method for the mass culture of ctenophores and other pelagic marine invertebrates. *BioRxiv* (doi:10.1101/751099)
193. Moroz, L. L. et al. 2014 The ctenophore genome and the evolutionary origins of neural systems. *Nature* **510**, 109–114. (doi:10.1038/nature13400)
194. Ryan, J. F. et al. 2013 The genome of the ctenophore *Mnemiopsis leidyi* and its implications for cell type evolution. *Science*

- 342, 1242592. (doi:10.1126/science.1242592)
195. Ludeman, D. A., Farrar, N., Riesgo, A., Paps, J. & Leys, S. P. 2014 Evolutionary origins of sensation in metazoans: functional evidence for a new sensory organ in sponges. *BMC Evol. Biol.* **14**, 3. (doi:10.1186/1471-2148-14-3)
196. Hammel, J. U. & Nickel, M. 2014 A new flow-regulating cell type in the Demosponge *Tethya wilhelma* - functional cellular anatomy of a leuconoid canal system. *PLoS One* **9**, e113153. (doi:10.1371/journal.pone.0113153)
197. Schippers, K. J., Sipkema, D., Osinga, R., Smidt, H., Pomponi, S. A., Martens, D. E. & Wijffels, R. H. 2012 Cultivation of sponges, sponge cells and symbionts: achievements and future prospects. *Adv Mar Biol* **62**, 273–337. (doi:10.1016/B978-0-12-394283-8.00006-0)
198. Rivera, A. S. et al. 2011 RNA interference in marine and freshwater sponges: actin knockdown in *Tethya wilhelma* and *Ephydatia muelleri* by ingested dsRNA expressing bacteria. *BMC Biotechnol.* **11**, 67. (doi:10.1186/1472-6750-11-67)
199. Revilla-I-Domingo, R., Schmidt, C., Zifko, C. & Raible, F. 2018 Establishment of Transgenesis in the Demosponge *Suberites domuncula*. *Genetics* **210**, 435–443. (doi:10.1534/genetics.118.301121)
200. Srivastava, M. et al. 2010 The Amphimedon queenslandica genome and the evolution of animal complexity. *Nature* **466**, 720–726. (doi:10.1038/nature09201)
201. Ryu, T. et al. 2016 Hologenome analysis of two marine sponges with different microbiomes. *BMC Genomics* **17**, 158. (doi:10.1186/s12864-016-2501-0)
202. Booth, D. S., Szmidt-Middleton, H. & King, N. 2018 Choanoflagellate transfection illuminates their cell biology and the ancestry of animal septins. *Mol. Biol. Cell* , mbcE18080514. (doi:10.1091/mbc.E18-08-0514)
203. King, N. et al. 2008 The genome of the choanoflagellate *Monosiga brevicollis* and the origin of metazoans. *Nature* **451**, 783–788. (doi:10.1038/nature06617)
204. Fairclough, S. R. et al. 2013 Premetazoan genome evolution and the regulation of cell differentiation in the choanoflagellate *Salpingoeca rosetta*. *Genome Biol.* **14**, R15. (doi:10.1186/gb-2013-14-2-r15)
205. Richter, D. J., Fozouni, P., Eisen, M. B. & King, N. 2018 Gene family innovation, conservation and loss on the animal stem lineage. *Elife* **7**. (doi:10.7554/eLife.34226)