

**Mass estimation of extinct taxa
and phylogenetic data both influence analyses of character
evolution in a large clade of birds (Telluraves)
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S1 COMPETITIVE RELEASE FOR TELLURAVES SPECIES FOLLOWING THE K-PG BOUNDARY

In the analyses of unmodified phylogenetic data and mass estimates derived from all available data we recovered support for a BM_{shift} model where the rate of evolution (σ^2) increased from 0.003 to 0.027. This increase in rate is potentially suggestive of competitive release as differentiation in body size linked to differences in resource use (Lack, 1946; Ashmole, 1968; Schoener, 1974; Wilson, 1975; Santa Rosalia reconsidered: size ratios and competition, 1981; Werner and Gilliam, 1984; Martin, 1985). During the Cretaceous ornithurines (all modern birds and their nearest relatives) likely faced significant competitive pressure from the now extinct enantiornithines (Dyke and Nudds, 2008), and perhaps even pterosaurs (Butler et al., 2009; Benson et al., 2014). Competition was likely strongest with enantiornithines because their skeletal elements, especially the forelimbs, had similar proportions and thus likely served similar functions (Dyke and Nudds, 2008). Additionally, enantiornithines were a well-established, diverse group before ornithurines are hypothesized to have originated, so they might have suppressed early ornithurine diversity. Extinction of competing groups at the K-Pg extinction event likely created many ecological opportunities for neornithines (modern birds, in which Telluraves forms a monophyletic group) to radiate, as has been frequently suggested (Jarvis et al., 2014; Prum et al., 2015). Although this ecological opportunity would have been sudden, our results suggest only gradual increase in variation through the Cenozoic. No burst of size variation early in the clade's history may be due to differentiation with respect to morphological shape, rather than size. Alternatively, this result may be explained by ecological constraint. Extant members of Telluraves are principally arboreal foragers, so their need to maneuver and perch in close environments places additional restrictions on body size on top of the physiological constraints experienced by all species. Although some large species can forage in trees (for example, the Great hornbill (*Buceros bicornis*) can weigh up to 4kg; Dunning, 2007), they are uncommon. Only two extinct species fall outside of the range of the extant species included here (*Messelirrisor halcyrostris* and *M. parvus*, Mayr 1998ax), which may suggest that extinct species experienced similar constraints, but this hypothesis would benefit from further investigation.

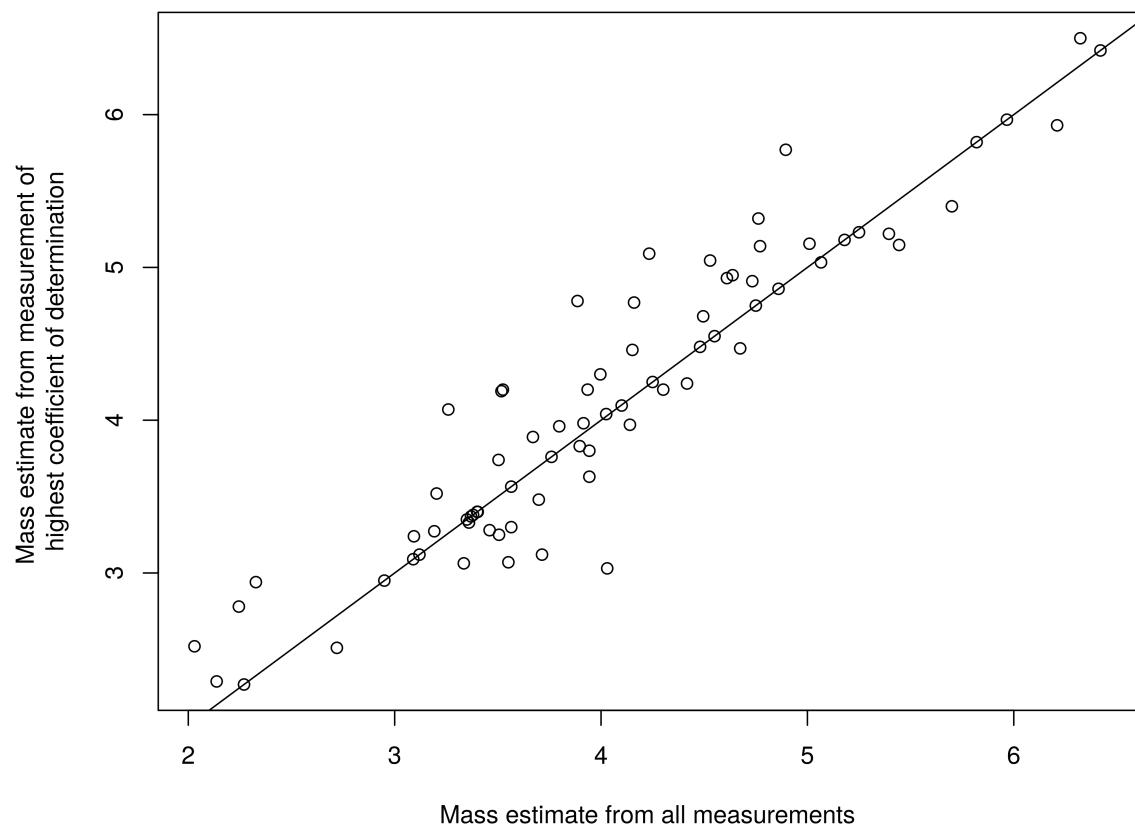


Figure S1: Comparison of mass estimates for 76 extinct species between using all available measurements and only using the measurement with the highest coefficient of determination. The solid lines denotes a 1 to 1 relationship.

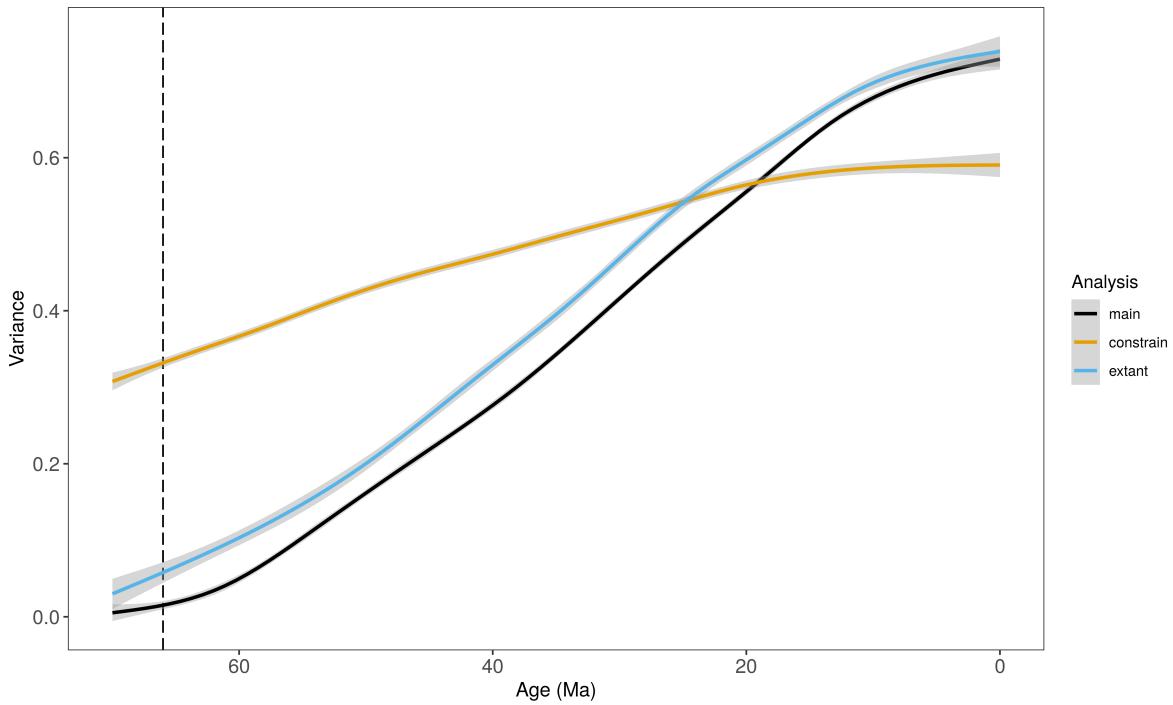


Figure S2: Maximum variance through time based on three models of evolution, when all available measurements were used to estimate the mass of extinct taxa. Main: the principal phylogenetic result from Crouch et al. (2019); constrain: phylogenetic trees with topological constraints to match current hypotheses regarding species relationships; and extant: analysis of only extant taxa. Prediction intervals in gray show variation in estimates due to topological variation in the sampled phylogenetic data.

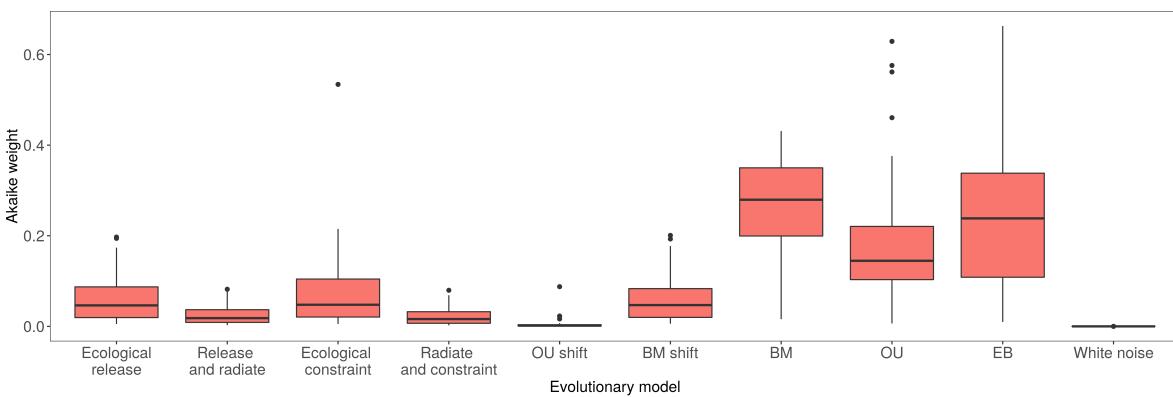


Figure S3: Distribution of Akaike weight scores for the ten models of evolution fit only to the analysis of extant taxa.

Table S1: Extinct taxa included in this study. References denoted by superscript numbers are provided in Table S2. The mean and standard errors for each species is provided in a separate .csv file.

Species	Age (Ma)	Element	Measurement(mm)	ln(Estimated Body Size)
<i>Nelepsittacus daphneleae</i>	16 - 19 ¹	Tarsometatarsus Shaft Width ⁴³	4.0	6.42
<i>Nelepsittacus donmertoni</i>	16 - 19 ¹	Humerus Shaft Width ⁴³	3.3	4.96
		Humerus Shaft Width ⁴³	3.3	4.96
		Humerus Shaft Width ⁴³	3.6	5.18
		Humerus Shaft Width ⁴³	3.4	5.03
		Tarsometatarsus Shaft Width ⁴³	2.4	5.20
<i>Nelepsittacus minimus</i>	16 - 19 ¹	Tarsometatarsus Shaft Width ⁴³	2.0	4.77
		Tarsometatarsus Length ⁴³	17.8	3.55
<i>Australopicus nelsonmandelai</i>	3.6 - 5.3 ²	Tarsometatarsus Shaft Width ²	2.2	5.00
		Tarsometatarsus Shaft Width ²	2.1	4.89
		Tarsometatarsus Shaft Width ²	2.1	4.89
		Tarsometatarsus Shaft Width ²	2.0	4.77
		Tarsometatarsus Shaft Width ²	2.3	5.10
		Tarsometatarsus Shaft Width ²	2.3	5.10
		Tarsometatarsus Length ²	24.3	4.24
		Tarsometatarsus Length ²	24.1	4.23
		Tarsometatarsus Length ²	23.1	4.13
		Tarsometatarsus Length ²	22.8	4.10
		Tarsometatarsus Length ²	25.0	4.31
<i>Piculoides saulcetensis</i>	20.4 - 23 ⁴	Tarsometatarsus Shaft Width ⁴	1.1	3.35
<i>Ecoracias brachyptera</i>	40.4 - 48.6 ¹	Femur Length ³¹	32.1	5.04
		Tibiotarsus Length ³¹	41.3	4.26
		Tarsometatarsus Length ³¹	18.2	3.60
		Tarsometatarsus Length ³¹	18.2	3.60
		Tibiotarsus Length ³¹	36.4	3.92
		Tibiotarsus Length ³¹	36.2	3.91
		Tarsometatarsus Length ³¹	18.1	3.59
		Humerus Length ³¹	44.4	5.08
		Humerus Length ³¹	44.7	5.10
		Humerus Length ³¹	47.7	5.23
<i>Geranopterus alatus</i>	33.9 - 37.2 ¹	Coracoid Shaft Width ³¹	2.7	5.27
<i>Geranopterus milneedwardsi</i>	33.9 - 37.2 ¹	Humerus Width ³¹	3.6	5.18
<i>Pici</i> indet	23 - 28.4 ⁵	Tarsometatarsus Shaft Width ⁵	1.0	3.12
<i>Primozygodactylus major</i>	40.4 - 48.6 ⁶	Tarsometatarsus Length ⁶	28.0	4.56
		Tibiotarsus Length ⁶	39.0	4.11
		Tibiotarsus Length ⁶	39.8	4.16
		Tibiotarsus Length ⁶	42.8	4.36
		Tibiotarsus Length ⁶	44.0	4.43
		Femur Length ⁶	24.6	4.29
		Humerus Length ⁶	29.0	4.20
		Femur Length ⁶	17.5	3.33
<i>Primozygodactylus eunjoiae</i>	40.4 - 48.6 ⁶	Tibiotarsus Length ⁶	29.8	3.39
<i>Zygodactylus luberonensis</i>	28.4 - 33.9 ¹	Humerus Length ⁶	17.2	3.12
		Humerus Length ⁶	17.2	3.12
		Tibiotarsus Length ⁶	34.8	3.80
		Tibiotarsus Length ⁶	34.7	3.80
		Tarsometatarsus Length ⁶	24.6	4.27
		Tarsometatarsus Length ⁶	24.5	4.26
<i>Primozygodactylus danielsi</i>	48 - 51 ⁷	Humerus Length ⁶	16.4	3.02
		Humerus Length ⁶	16.5	3.03
		Humerus Length ⁶	17.4	3.14
		Femur Length ⁶	16.5	3.17
		Femur Length ⁶	17.2	3.28
		Femur Length ⁶	16.0	3.08
		Tibiotarsus Length ⁶	27.4	3.16
		Tibiotarsus Length ⁶	27.3	3.15
		Tibiotarsus Length ⁶	29.4	3.35

Species	Age (Ma)	Element	Measurement(mm)	ln(Estimated Body Size)
Primozygodactylus ballmanni	48 - 51 ⁷	Tibiotarsus Length ⁶	27.5	3.17
		Tarsometatarsus Length ⁶	19.6	3.77
		Tarsometatarsus Length ⁶	19.7	3.78
		Tarsometatarsus Length ⁶	20.7	3.89
		Tarsometatarsus Length ⁶	19.0	3.70
Eobucco brodkorbi	39 - 50 ¹	Humerus Length ⁶	21.0	3.53
Neanis kistneri	50.3 - 55.8 ²⁹	Humerus Length ⁶	20.5	3.48
Eoglaucidium pallas	48.5 - 55.8 ¹⁰	Humerus Length ⁶	20.0	3.43
		Femur Length ⁶	20.8	3.82
		Tibiotarsus Length ⁶	33.0	3.66
		Tarsometatarsus Length ⁶	24.6	4.27
		Tarsometatarsus Length ²⁹	26.95	4.48
		Humerus Shaft Width ²⁹	1.8	3.4
		Humerus Length ¹⁰	46.3	5.17
		Humerus Length ¹⁰	43.9	5.06
		Humerus Length ¹⁰	47.9	5.24
		Humerus Length ¹⁰	44.1	5.07
		Humerus Length ¹⁰	46.6	5.18
		Humerus Length ¹⁰	44.2	5.07
		Humerus Length ¹⁰	46.6	5.18
		Femur Length ¹⁰	35.9	5.36
		Femur Length ¹⁰	31.9	5.02
		Tibiotarsus Length ¹⁰	45.7	4.53
		Tibiotarsus Length ¹⁰	46.5	4.58
		Tibiotarsus Length ¹⁰	49.5	4.75
		Tibiotarsus Length ¹⁰	49.6	4.75
		Tarsometatarsus Length ¹⁰	22.2	4.04
		Tarsometatarsus Length ¹⁰	22.2	4.04
		Tarsometatarsus Length ¹⁰	22.3	4.05
		Tarsometatarsus Length ¹⁰	26.0	4.40
		Tarsometatarsus Length ¹⁰	25.8	4.38
		Tarsometatarsus Length ¹⁰	24.2	4.24
Eoglaucidium spp	48.5 - 55.8 ¹⁰	Tibiotarsus Length ¹⁰	35.7	3.87
		Tarsometatarsus Length ¹⁰	22.2	4.04
		Tarsometatarsus Length ¹⁰	21.1	3.93
		Femur Length ¹⁰	25.0	4.34
		Femur Length ¹⁰	27.2	4.58
		Humerus Length ¹⁰	19.7	3.40
Masillacolius brevidactylus	46 - 52 ¹⁰	Tibiotarsus Length ¹⁰	21.5	2.51
		Tarsometatarsus Length ¹⁰	20.4	3.85
		Tarsometatarsus Length ¹⁰	20.4	3.85
Anneavis anneae	50.3 - 55.4 ¹⁵	Coracoid Length ¹⁵	23.2	4.51
		Coracoid Length ¹⁵	22.8	4.46
		Humerus Length ¹⁵	41.7	4.95
		Humerus Length ¹⁵	40.1	4.87
		Femur Length ¹⁵	27.7	4.63
		Femur Length ¹⁵	33.4	5.15
		Femur Diameter ¹⁵	2.8	5.20
		Femur Diameter ¹⁵	2.8	5.20
		Tarsometatarsus Length ¹⁵	23.7	4.19
		Tarsometatarsus Length ¹⁵	23.5	4.17
Sandcoleus copiosus	50.3 - 55.8 ¹⁵	Coracoid Length ¹⁵	24.7	4.70
		Coracoid Length ¹⁵	24.4	4.67
		Humerus Length ¹⁵	46.3	5.17
		Humerus Length ¹⁵	45.6	5.14
		Femur Length ¹⁵	36.7	5.42
		Femur Length ¹⁵	36.5	5.40
		Tarsometatarsus Length ¹⁵	28.1	4.57
Chascacolius oscitans	50 - 54 ¹⁵	Humerus Length ¹⁵	26.5	4.01
		Coracoid Length ¹⁵	17.8	3.70
		Humerus Length ¹⁵	26.6	4.04
		Femur Length ¹⁵	25.1	4.35
Rupelramphastoides knopfi	28.4 - 33.9 ¹	Humerus Length ⁴⁵	11.5	2.29
		Femur Length ⁴⁵	10.8	1.97

Species	Age (Ma)	Element	Measurement(mm)	ln(Estimated Body Size)
Serudaptus pohli	40.4 - 48.6 ⁸	Tibiotarsus Length ⁴⁵	19.0	2.18
		Tibiotarsus Length ⁴⁵	18.5	2.11
		Humerus Length ⁸	38.4	4.78
		Tibiotarsus Length ⁸	38.2	4.05
		Tibiotarsus Length ⁸	37.4	4.00
		Tarsometatarsus Length ⁸	15.7	3.27
Sandcoleidae indet	45 - 52 ⁸	Tarsometatarsus Length ⁸	16.1	3.33
		Humerus Length ⁸	40.8	4.91
		Humerus Length ⁸	41.6	4.95
		Femur Length ⁸	32.6	5.09
		Tibiotarsus Length ⁸	42.4	4.33
		Tarsometatarsus Length ⁸	23.7	4.19
Avolatavis tenens	51.57 - 51.75 ²²	Tarsometatarsus Length ⁸	23.7	4.19
		Femur Length ²²	24.7	4.30
		Tibiotarsus Length ²²	40.8	4.23
		Tarsometatarsus Length ²²	17.1	3.46
		Tibiotarsus Length ¹⁶	21.5	2.51
		Tarsometatarsus Length ¹⁶	13.5	2.93
Zygodactylidae UWGM21421	50.3 - 55.8 ¹⁶	Coracoid Length ¹⁶	13.8	2.92
		Humerus Length ¹⁶	16.8	3.07
		Humerus Length ¹⁶	16.8	3.07
		Femur Length ¹⁶	19.7	3.67
		Tibiotarsus Length ¹⁶	30.6	3.46
		Tarsometatarsus Length ¹⁶	21.7	3.99
Eozygodactylus americanus	50.3 - 55.8 ¹⁷	Tarsometatarsus Length ¹⁶	21.8	4.00
		Tarsometatarsus Length ¹⁶	20.9	3.91
		Tarsometatarsus Length ¹⁶	20.5	3.87
		Coracoid ¹⁶	13.7	2.90
		Humerus Length ¹⁶	18.3	3.25
		Femur Length ¹⁶	16.8	3.22
Zygodactylus grandis	50.3 - 55.8 ¹⁶	Tibiotarsus Length ¹⁶	33.0	3.66
		Tibiotarsus Length ¹⁶	33.6	3.71
		Tarsometatarsus Length ¹⁶	21.4	3.95
		Tarsometatarsus Length ¹⁶	20.2	3.83
		Coracoid ⁹	19.8	4.03
		Coracoid ⁹	21.6	4.29
Messelirrisor parvus	40.4 - 48.6 ¹	Tarsometatarsus Length ⁹	17.95	3.57
		Tarsometatarsus Diameter ⁹	1.35	3.83
		Coracoid ⁵¹	8.8	1.54
		Humerus Length ⁵¹	12.9	2.52
		Coracoid ⁵¹	9.3	1.71
		Humerus Length ⁵¹	14.6	2.78
Messelirrisor halcyrostris	40.4 - 48.6 ¹	Humerus Length ⁵¹	19.3	3.36
		Humerus Length ⁵¹	20.1	3.44
		Humerus Length ¹⁹	49.8	5.32
		Femur Length ¹⁹	29.0	4.76
		Tibiotarsus Length ¹⁹	46.2	4.56
		Tarsometatarsus Length ¹⁹	26.2	4.41
Messelirrisor grandis	40.4 - 48.6 ¹	Tarsometatarsus Length ¹⁹	30.0	4.71
		Tarsometatarsus Length ¹⁹	32.0	4.86
		Tarsometatarsus Length ¹⁹	34.2	5.01
		Tibiotarsus Length ¹⁹	79.1	6.00
		Tarsometatarsus Length ¹⁹	40.8	5.40
		Coracoid ¹⁹	33.7	5.65
Plesiocathartes kelleri	51 - 54.5 ¹³	Coracoid ¹⁹	34.7	5.74
		Tibiotarsus Length ¹⁹	69.1	5.64
		Tibiotarsus Length ¹⁹	69.1	5.64
		Tarsometatarsus Length ¹⁹	36.4	5.15
		Tarsometatarsus Length ¹⁹	36.6	5.16
		Tarsometatarsus Length ¹⁹	36.2	5.13
Plesiocathartes europaeus	33 - 45 ¹⁸	Humerus Length ¹⁹	62.0	5.77
		Tarsometatarsus Length ¹⁹	26.0	4.40
		Tarsometatarsus Length ¹⁹	27.0	4.48
		Tibiotarsus Length ¹⁹	53.0	4.93
Plesiocathartes geiselensis	33 - 45 ¹⁸			
Plesiocathartes major	33 - 45 ¹⁹			
Plesiocathartes wyomingensis	33 - 45 ¹⁹			

Species	Age (Ma)	Element	Measurement(mm)	ln(Estimated Body Size)
<i>Oligocolius brevitarsus</i>	28.4 - 33.9 ¹	Femur Length ⁴⁶	21.6	3.92
		Tibiotarsus Length ⁴⁶	27.6	3.18
<i>Vastanavis cambayensis</i>	48.6 - 55.8 ¹	Tarsometatarsus Length ⁴⁶	15.8	3.28
		Coracoid Width ⁴⁹	3.3	5.73
<i>Vastanavis eocaena</i>	48.6 - 55.8 ¹	Coracoid Width ⁴⁹	3.4	5.80
		Coracoid Width ⁴⁹	3.6	5.93
<i>Messelastur gratulator</i>	40.4 - 48.6 ¹	Coracoid Width ⁴⁹	3.9	6.11
		Coracoid Width ⁴⁹	3.6	5.93
<i>Psittacopes lepidus</i>	48 - 52 ¹¹	Coracoid Width ⁴⁹	3.5	5.86
		Humerus Length ⁴⁷	36.6	4.68
<i>Pulchrapolia gracilis</i>	48.6 - 55. 8 ¹²	Tibiotarsus Length ⁴⁷	42.0	4.31
		Humerus Length ¹¹	18.3	3.25
<i>Eocolius walkeri</i>	54.4 - 56.4 ²⁰	Humerus Length ¹¹	18.2	3.24
		Humerus Length ¹¹	18.8	3.30
<i>Palaeospiza bella</i>	33.9 - 37.2 ¹	Humerus Length ¹¹	18.8	3.30
		Tarsometatarsus Length ¹¹	13.1	2.87
<i>Primocolius minor</i>	37.2 - 40.4 ¹	Femur Length ¹²	20.6	3.79
		Femur width ¹²	1.7	3.96
<i>Primocolius sigei</i>	37.2 - 40.4 ¹	Coracoid width ¹²	1.6	4.09
		Tarsometatarsus Length ¹²	16.3	3.35
<i>Oligocolius psittacocephalon</i>	24.6 - 24.8 ²¹	Coracoid ²⁰	18.4	3.80
		Humerus Length ²⁰	22.0	3.63
<i>Primotrogon wintersteini</i>	28.4 - 33.9 ¹	Tarsometatarsus Length ²⁰	26.0	4.40
		Humerus Length ⁴⁸	23.2	3.74
<i>Quasisyndactylus longibrachis</i>	45 - 55 ²³	Tibiotarsus Length ⁴⁸	28.1	3.23
		Tarsometatarsus Length ⁴⁸	17.7	3.54
<i>Phirriculus pinicola</i>	17 - 22 ²⁴	Tarsometatarsus Length ²¹	16.4	3.37
		Humerus Length ²¹	23.5	3.76
<i>Paleotodus itardiensis</i>	28.4 - 33.9 ²⁷	Humerus Length ²¹	25.0	3.89
		Tarsometatarsus Length ²¹	17.0	3.45
<i>Paleotodus emryi</i>	33.3 - 33.9 ²⁸	Tarsometatarsus Length ⁵⁰	12.7	2.80
		Tibiotarsus Length ⁵⁰	24.9	2.91
<i>Selmes absurdipes</i>	42 - 48 ³⁰	Humerus Length ⁵⁰	27.2	4.07
		Humerus Length ²³	15.8	2.94
<i>Upupa antaios</i>	0.0001 - 0.001 ³³	Tibiotarsus Length ²³	17.3	1.93
		Tarsometatarsus Length ²³	9.8	2.22
<i>Paracoracias occidentalis</i>	45 - 55 ³⁵	Tarsometatarsus Length ²³	9.8	2.22
		Humerus Length ²⁴	21.6	3.59
<i>Mopsitta tanta</i>	48.6 - 55.8 ¹	Humerus Length ²⁴	21.2	3.54
		Femur Length ²⁷	14.7	2.84
		Coracoid ²⁷	15.1	3.20
		Humerus Length ²⁷	18.2	3.24
		Femur Length ²⁸	16.6	3.18
		Humerus Length ²⁸	20.9	3.52
		Tibiotarsus Length ²⁸	24.9	2.91
		Tarsometatarsus Length ³⁰	20.2	3.83
		Tibiotarsus Length ³⁰	28.8	3.30
		Tarsometatarsus Length ³¹	30.5	4.75
		Humerus Length ³³	33.1	4.47
		Humerus Width ³³	3.1	4.80
		Humerus Width ³³	3.1	4.80
		Humerus Width ³³	3.2	4.88
		Femur Length ³³	28.1	4.67
		Femur Width ³³	2.5	4.92
		Tibiotarsus Length ³³	40.1	4.18
		Femur Length ³⁵	29.3	4.78
		Femur Length ³⁵	29.5	4.80
		Humerus Length ³⁵	43.7	5.05
		Humerus Length ³⁵	43.6	5.04
		Tibiotarsus Length ³⁵	39.8	4.16
		Tibiotarsus Length ³⁵	39.6	4.15
		Tarsometatarsus Length ³⁵	19.2	3.72
		Humerus Length ³⁸	67.0	5.93
		Humerus Width ³⁸	6.0	6.49

Species	Age (Ma)	Element	Measurement(mm)	ln(Estimated Body Size)
<i>Psuedasturidae</i> indet	48.6 - 55.8 ³⁸	Humerus Length ³⁸	29	4.20
		Humerus Width ³⁸	2.0	3.67
<i>Bavaripsitta ballmanni</i>	13.5 - 15 ³⁹	Tarsometatarsus Length ³⁹	13.6	2.95
<i>Eclectus infectus</i>	0.01 - 1 ⁴²	Femur Length ⁴²	51.3	6.36
		Femur Width ⁴²	4.7	6.50
		Tarsometatarsus Width ⁴²	3.9	6.36
		Humerus Width ⁴²	5.1	6.07
<i>Primobucco perneri</i>	40.4 - 48.6 ¹	Humerus Length ⁵²	28.6	4.17
		Humerus Length ⁵²	28.9	4.19
		Humerus Length ⁵²	25.2	3.91
		Humerus Length ⁵²	26.5	4.01
		Humerus Length ⁵²	25.8	3.96
		Humerus Length ⁵²	29.3	4.22
		Humerus Length ⁵²	29.3	4.21
		Tarsometatarsus Length ⁵²	23.1	4.13
<i>Primobucco mcgrewi</i>	48.6 - 55.8 ¹	Tibiotarsus length ⁵²	26.7	3.09
<i>Picavus litencicensis</i>	29.5 - 32 ⁵³	Femur Length ⁵³	12	2.27
<i>Pseudastur macrocephalus</i>	48 - 52 ⁵⁴	Humerus Length ⁵⁵	27.6	4.10
		Humerus Length ⁵⁵	30.5	4.30
		Humerus Length ⁵⁵	29.0	4.20
		Tibiotarsus Length ⁵⁵	30.5	3.45
		Tibiotarsus Length ⁵⁵	30.6	3.46
		Tibiotarsus Length ⁵⁵	31.9	3.57
		Tibiotarsus Length ⁵⁵	29.0	3.31
		Tarsometatarsus Length ⁵⁵	15.5	3.24
		Tarsometatarsus Length ⁵⁵	15.5	3.24
		Tarsometatarsus Length ⁵⁵	14.2	3.05
		Tarsometatarsus Length ⁵⁵	13.0	2.85
<i>Xenopsitta fejvari</i>	16 - 23 ⁵⁶	Tarsometatarsus Length ⁵⁶	16.5	3.38
<i>Capitonides protractus</i>	13.8 - 16 ⁴¹	Humerus Length ⁴¹	26.0	3.97
<i>Palaeopsittacus georgei</i>	48.6 - 55.8 ⁴⁰	Tarsometatarsus Length ⁴¹	25.0	4.31
<i>Brachypteryx langrandi</i>	0.001 - 0.002 ³²	Coracoid ⁴⁰	23.5	4.55
<i>Coliiformes</i> indet	45 - 52 ¹⁰	Humerus Length ³²	47.4	5.22
		Humerus Width ³²	4.2	5.57
<i>London Clay Species A</i>	48.6 - 55.8 ¹¹	Tarsometatarsus Length ¹⁰	20.4	3.85
		Humerus Length ¹⁰	25.8	3.98
		Coracoid ¹¹	21.2	4.24
		Tibiotarsus Length ¹¹	36	3.89
<i>Clericolius acriala</i>	51.57 - 51.75 ²¹	Tarsometatarsus Length ¹¹	19.1	5.12
		Tibiotarsus Length ²¹	34.8	3.80
		Tarsometatarsus Length ²¹	21.9	4.01
		Tarsometatarsus Length ²¹	22.0	4.02
<i>Motmot</i> indet	4.9 - 10.3 ²⁶	Humerus Shaft Width ²⁶	2.5	4.25
<i>Cyrilavis olsoni</i>	48.6 - 55.8 ¹	Humerus Length ⁴⁴	28.8	4.19
		Coracoid ⁴⁴	16.0	3.37
		Tibiotarsus Length ⁴⁴	28.5	3.27
		Tarsometatarsus Length ⁴⁴	15.5	3.24

Table S2: Associated references for the data presented in Table S1

Number	Reference
1	paleo-db.org
2	Manegold and Louchart (2012), horizon age checked on paleodb.org
3	Brodkorb (1970), horizon age checked on paleodb.org
4	De Pietri et al. (2011)
5	Mayr (2001b), horizon age checked on paleodb.org
6	Mayr and Zelenkov (2009), horizon age checked on paleodb.org
7	Mayr (1998c), horizon age checked on paleodb.org
8	Mayr (2000b), horizon age checked on paleodb.org
9	Mourer-Chauviré (1992), horizon age checked on paleodb.org
10	Mayr and Peters (1998), horizon age checked on paleodb.org
11	Mayr and Daniels (1998), horizon age checked on paleodb.org
12	Dyke and Cooper (2000), horizon age checked on paleodb.org
13	Mayr (2002), horizon age checked on paleodb.org
14	Boles (1997), horizon age checked on paleodb.org
15	Houde and Olson (1992), horizon age checked on paleodb.org
16	DeBee (2012), horizon age checked on paleodb.org
17	Weidig (2010), horizon age checked on paleodb.org
18	Weidig (2006), horizon age checked on paleodb.org
19	Weidig (2006)
20	Dyke and Waterhouse (2001)
21	Mayr (2013)
22	Ksepka and Clarke (2012)
23	Mayr (2004), horizon age checked on paleodb.org
24	Mlíkovský and Göhlich (2000), horizon age checked on paleodb.org
25	Mayr and Smith (2013)
26	Becker (1986), horizon age checked on paleodb.org
27	Mayr and Micklich (2010), horizon age checked on paleodb.org
28	Mayr and Knopf (2007)
29	Feduccia and Martin (1976), horizon age checked on paleodb.org
30	Mayr (2001a)
31	Mayr and Mourer-Chauviré (2000), horizon age checked on paleodb.org
32	Goodman (2010)
33	Olson (1975)
34	Mourer-Chauviré et al. (2013), horizon age checked on paleo-db
35	Clarke et al. (2009), horizon age checked on paleodb.org
36	Ksepka and Clarke (2010)
37	Mayr (2010), horizon age checked on paleodb.org
38	Waterhouse et al. (2008), same formation as <i>Mopsitta tanta</i>
39	Mayr and Göhlich (2004), horizon age checked on paleodb.org
40	Harrison (1982), horizon age checked on paleodb.org
41	Ballman (1983), horizon age checked on paleodb.org
42	Steadman (2006)
43	Worthy et al. (2011)
44	Ksepka et al. (2011)
45	Mayr (2005a)
46	Mayr (2000a)
47	Mayr (2005c)
48	Ksepka and Clarke (2009)
49	Mayr et al. (2010)

Number	Reference
50	Mayr (2005b)
51	Mayr (2000c)
52	Mayr et al. (2004)
53	Mayr and Gregorová (2012)
54	Morlo et al. (2004)
55	Mayr (1998b)
56	Mlíkovský (1998)

Table S3: Extant species included in this study. n is the total number of taxa represented by the species in the phylogeny. Note that this is not necessarily the total number of described species, but the number for which body mass data were available.

Order	Species in phylogeny	n	Additional taxa represented
Bucerotiformes	<i>Rhinopomastus cyanomelas</i>	3	Rhinopomastus
	<i>Upupa epops</i>	2	Upupa
	<i>Phoeniculus purpureus</i>	5	Phoeniculus
	<i>Tockus nasutus</i>	14	Tockus, Lophoceros
	<i>Rhyticeros undulatus</i>	33	Penelopides, Rhabdotorrhinus, Rhynceros, Aceros, Antrhacoceros, Ocyeros, Annorhinus, Buceros, Rhinoplax, Bycanistes, Ceratogymna, Tropicranus, Horizocerus, Berenicornis
Coliiformes	<i>Bucorvus abyssinicus</i>	2	<i>Bucorvus leadbeateri</i>
	<i>Colius colius</i>	2	<i>Colius castanotus</i>
	<i>Colius leucocephalus</i>	1	–
	<i>Colius striatus</i>	1	–
	<i>Urocolius indicus</i>	1	–
Trogoniformes	<i>Urocolius macrourus</i>	1	–
	<i>Trogon melanurus</i>	19	Trogon, Priotelus
	<i>Pharomachrus antisianus</i>	6	Pharomachrus, Euptilotis
Leptosomiformes	<i>Harpactes ardens</i>	11	Harpactes, Apaloderma, Apalharpactes
	<i>Leptosomus discolor</i>	1	–
	<i>Todus angustirostris</i>	2	<i>Todus multicolor</i>
	<i>Todus todus</i>	2	<i>Todus mexicanus</i>
	<i>Todus subulatus</i>	1	–
Coraciiformes	<i>Coracias garrulus</i>	8	Coracias
	<i>Eurystomus orientalis</i>	4	Eurystomus
	<i>Atelornis corsleyi</i>	1	–
	<i>Atelornis pittoides</i>	1	–
	<i>Brachypteracias leptosomus</i>	1	–
Piciformes	<i>Brachypteracias squamigera</i>	1	–
	<i>Alcedo atthis</i>	2	<i>Alcedo coerulescens</i>
	<i>Alcedo quadribrachys</i>	2	<i>Alcedo semitorquata</i>
	<i>Alcedo meninting</i>	2	<i>Alcedo hercules, Alcedo euryzona</i>
	<i>Alcedo leucogaster</i>	6	Corythornis, Ispidina
	<i>Ceyx pictus</i>	6	Ceyx
	<i>Megacyrle alcyon</i>	4	Megacyrle
	<i>Chloroceryle americana</i>	5	Chloroceryle, Ceryle
	<i>Todiramphus chloris</i>	16	Halcyoninae
	<i>Merops orientalis</i>	25	Meropidae
	<i>Hylomanes momotula</i>	4	Eumomota, Electron
	<i>Momotus mexicanus</i>	6	Momotus, Baryphthengus, Aspatha
	<i>Malacoptila semicincta</i>	31	Bucconidae
	<i>Galbulia ruficauda</i>	16	Galbulidae
	<i>Indicator maculatus</i>	17	Indicatoridae
	<i>Lybius dubius</i>	7	Lybius
	<i>Pogoniulus bilineatus</i>	9	Pogoniulus
	<i>Trachyphonus darnaudii</i>	13	Trachyphonus, Cryptolybia, Buccanodon, Gymnobucco, Stactolaema
	<i>Megalaima virens</i>	19	Megalaimidae

Order	Species in phylogeny	n	Additional taxa represented
	<i>Picumnus cirratus</i>	27	Jynx, Sasia, Picumnus, Nesocites, Hemicircus
	<i>Blythipicus rubiginosus</i>	2	<i>Blythipicus pyrrhotis</i>
	<i>Chrysocolaptes lucidus</i>	3	Reinwardtipicus, Chrysocolaptes
	<i>Campephilus haematogaster</i>	7	Campephilus
	<i>Campethera caroli</i>	73	Picini
	<i>Dendrocopos major</i>	86	Melanerpini
	<i>Ramphastos toco</i>	7	Ramphastos
	<i>Pteroglossus torquatus</i>	13	Pteroglossus
	<i>Andigena cucullata</i>	5	Andigena, "Selenidera" spectabilis
	<i>Selenidera maculirostris</i>	5	Selenidera
	<i>Aulacorhynchus derbianus</i>	6	Aulacorhynchus
	<i>Capito niger</i>	13	Capitonidae
	<i>Semnornis ramphastinus</i>	2	Semnornis
Psittaciformes	<i>Cacatua galerita</i>	21	Cacatuidae
	<i>Poicephalus gulielmi</i>	8	Poicephalus, Psittacus
	<i>Bolborhynchus lineola</i>	10	Amoropsittaci
	<i>Brotogeris chiriri</i>	8	Brotogeris
	<i>Trichilaria malachitacea</i>	10	Pionopsitta, Trichilaria, Hapalopsittaca, Pyrilia, Graydascalus, Alipiopsitta
	<i>Pionus maximiliani</i>	4	Pionus
	<i>Barnardius zonarius</i>	1	–
	<i>Myiopsitta monachus</i>	1	–
	<i>Nestor notabilis</i>	2	Nestor
	<i>Strigops habroptilus</i>	1	–
	<i>Amazona amazonica</i>	25	Amazona
	<i>Phyrrhura picta</i>	52	Arini
	<i>Coracopsis vasa</i>	4	Psittrichasinae
	<i>Psittacula krameri</i>	32	Psittaculini
	<i>Psittacella brehmii</i>	4	Psittacella
	<i>Northiella haematogaster</i>	1	–
	<i>Pezoporus wallicus</i>	1	–
	<i>Psephotus haematonotus</i>	1	–
	<i>Neophema splendida</i>	7	Neosephotus, Neophema
	<i>Purpleicephalus spurius</i>	5	Psephotellus
	<i>Platycercus eximius</i>	6	Platycercus
	<i>Lathamus discolor</i>	2	Pyrrhulopsis
	<i>Cyanoramphus novaezelandiae</i>	5	Eunymphicus, Cyanoramphus
	<i>Agapornis roseicollis</i>	15	Agapornithini
	<i>Trichoglossus haematocephalus</i>	37	Loriini
Passeriformes	<i>Ancanthisittca chloris</i>	3	Xenicus
	<i>Pitta versicolor</i>	43	Eurylaimides
	<i>Tyrannus tyrannus</i>	288	Tyrannidae
	<i>Thamnophilus ruficapillus</i>	188	Thamnophilidae
	<i>Conopophaga lineata</i>	14	Conopophagidae, Melanopareiidae
	<i>Grallaria ruficapilla</i>	41	Grallariidae
	<i>Formicarius colma</i>	58	Rhinocryptidae, Formicariidae
	<i>Sclerurus rufigularis</i>	15	Sclerurinae
	<i>Dendrocincus merula</i>	15	Glyphorynchus, Certhiasomus, Sittasomus, Deconychura, Dendrocincus
	<i>Xiphocolaptes major</i>	18	Nasica, Dendrexetastes, Dendrocincus, Hylexetastes, Xiphocolaptes

Order	Species in phylogeny	n	Additional taxa represented
	<i>Xiphorhynchus fuscus</i>	24	Xiphorhynchus, Dendroplex, Campylorhamphus
	<i>Lepidocolaptes angustirostris</i>	9	Lepidocolaptes, Drymornis, Drymotoxeres
	<i>Pygarrhichas albogularis</i>	4	Pygarrhichadini, Berlepschiini
	<i>Lochmias nematura</i>	18	Lochmias, Geocerthia, Upucerthia, Cinclodes, Phleocryptes, Limnornis
	<i>Furnarius rufus</i>	6	Furnarius
	<i>Pseudocolaptes boissonneautii</i>	3	Pseudocolaptes, Tarphonomus, Premnornis
	<i>Philydor pyrrhodes</i>	20	Anabazenops, Megaxenops, Cichlocolaptes, Heliobletus, Philydor, Anabacerthia, Syndactyla
	<i>Automolus infuscatus</i>	16	Ancistros, Clibanornis, Thripadectes, Automolus
	<i>Margarornis rubiginosus</i>	4	Margarornini
	<i>Asthenes baeri</i>	38	Aphrastura, Leptasthenura, Phacellodomus, Hellmayrea, Coryphistera, Anumbius, Asthenes
	<i>Synallaxis frontalis</i>	27	Mazaria, Schoeniophylax, Certhiaxis, Synallaxis
	<i>Atrichornis clamosus</i>	2	Atrichornis rufescens
	<i>Menura novaehollandiae</i>	2	Menura alberti
	<i>Climacteris picumnus</i>	6	Climacteris
	<i>Cormobates leucophaeus</i>	2	Cormobates placens
	<i>Ailuroedus crassirostris</i>	3	Ailuroedus
	<i>Ptilonorhynchus violaceus</i>	6	Ptilonorhynchus, Chlamydera
	<i>Sericulus chrysocephalus</i>	4	Sericulus, Scenopoeetes, Amblyornis
	<i>Amytornis goyderi</i>	10	Amytornis
	<i>Malurus cyaneus</i>	23	Malurus, Chenorhamphus, Clytomyias, Sipodotus, Stipiturus
	<i>Dasyornis broadbenti</i>	3	Dasyornis
	<i>Pardalotus striatus</i>	7	Pardalotus
	<i>Sericornis frontalis</i>	77	Pachycare, Oreoscopus, Gerygone, Acanthornis, Aphelocephala, Acanthiza, Smicrornis, Pycnoptilus, Pyrrholaeus, Chthonicola, Hylacola, Calamanthus, Origma, Crateroscelis, Sericornis
	<i>Philemon citreogularis</i>	62	Phlidonyrini, Melithreptini, Philemonini, Myzomelini
	<i>Acanthorhynchus tenuirostris</i>	2	Acanthorhynchus
	<i>Certhionyx variegatus</i>	27	Prosthemaderini, Gliciphilini, Ephthianurini
	<i>Anthochaera carunculata</i>	53	Meliphaginae
	<i>Orthonyx temminckii</i>	3	Orthonyx
	<i>Cracticus torquatus</i>	6	Cracticus
	<i>Peltops montanus</i>	2	Peltops
	<i>Strepera graculina</i>	3	Strepera
	<i>Artamus cinereus</i>	11	Artamus
	<i>Pyrrhocorax pyrrhocorax</i>	49	Pyrrhocoracinae, Cissinae, Perisoreinae, Cyanococinae
	<i>Garrulus glandarius</i>	3	Garrulus
	<i>Pica pica</i>	9	Pica, Zavattariornis, Ptilostomus, Podoces
	<i>Nucifraga caryocatactes</i>	3	Nucifraga
	<i>Corvus corax</i>	38	Corvus, Coloeus
	<i>Parus major</i>	52	Paridae
	<i>Sitta frontalis</i>	19	Sittidae
	<i>Certhia familiaris</i>	7	Certhiidae
	<i>Thryothorus ludovicianus</i>	56	Thryothorinae

Order	Species in phylogeny	<i>n</i>	Additional taxa represented
	<i>Troglodytes troglodytes</i>	34	Odontorchilus, Catherpes, Hylorchilus, Salpinectes, Microcerculus, Nannus, Ferminia, Cistothorus, Thryophilus, Troglodytes
	<i>Sturnella neglecta</i>	10	Sturnella, Dolichonyx
	<i>Caicus melanicterus</i>	16	Cacicinae
	<i>Icterus spurius</i>	33	Icterus
	<i>Agelaius phoeniceus</i>	11	Agelaius, Nesopsar
	<i>Molothrus ater</i>	49	Agelaiinae
	<i>Rhodospiza obsoleta</i>	178	Fringillidae
	<i>Passer domesticus</i>	54	Passeridae
	<i>Mniotilla varia</i>	110	Parulidae
	<i>Calcarius lapponicus</i>	1	–
	<i>Calcarius pictus</i>	1	–
	<i>Plectrophenax hyperboreus</i>	1	–
	<i>Plectrophenax nivalis</i>	1	–
	<i>Xanthocephalus xanthocephalus</i>	1	–
	<i>Trichodroma muraria</i>	1	–

Table S4: AIC and AIC_w scores for model fitting analyses that include extinct taxa. Mean values are reported, with standard deviations in parentheses. ‘Main’ is where untransformed phylogenetic data were used in the analysis, ‘constraint’ is where phylogenetic data that had been reconstructed with topological constraints was used, and ‘scale’ is where phylogenetic data that had been scaled to have entirely Cenozoic divergence dates was used. For the scale analysis, only models describing a single mode of evolution were fit to the data. Dashes indicate where the fit of models was not estimated.

Morphological data	Model	Main analysis		Constraint		Scale	
		AIC	ΔAIC_w	AIC	ΔAIC_w	AIC	ΔAIC_w
<i>All available</i>							
Post-K-Pg release	4.16 (4.61)	0.13 (0.10)	3.04 (3.09)	0.13 (0.10)	—	—	—
Post-K-Pg release and radiate	6.07 (4.73)	0.05 (0.05)	3.83 (2.86)	0.09 (0.09)	—	—	—
Post-K-Pg constraint	5.09 (4.69)	0.10 (0.11)	4.30 (2.92)	0.08 (0.07)	—	—	—
Radiate, post-K-Pg constraint	3.21 (3.85)	0.18 (0.17)	5.25 (3.23)	0.07 (0.15)	—	—	—
OU	7.89 (4.60)	0.03 (0.05)	5.44 (4.04)	0.08 (0.12)	—	—	—
<i>OU shift</i>	3.84 (6.69)	0.35 (0.35)	3.38 (2.90)	0.23 (0.25)	—	—	—
BM	28.46 (40.70)	0.00 (0.00)	8.61 (8.79)	0.08 (0.14)	52.41 (29.50)	0.00 (0.00)	—
BM <i>shift</i>	10.61 (6.08)	0.02 (0.05)	6.86 (5.87)	0.06 (0.07)	12.54 (12.35)	0.23 (0.37)	—
OU	26.42 (41.75)	0.01 (0.02)	10.66 (8.79)	0.03 (0.05)	5.24 (12.16)	0.55 (0.39)	—
Early burst	24.34 (33.6)	0.02 (0.08)	7.75 (7.28)	0.06 (0.08)	10.13 (14.00)	0.15 (0.20)	—
BM with trend	62.21 (18.44)	0.02 (0.12)	101.59 (14.25)	0.00 (0.00)	34.04 (17.66)	0.06 (0.21)	—
White noise	—	—	—	—	—	—	—
<i>No intraspecific error</i>							
Post-K-Pg release	28.70 (24.69)	0.00 (0.01)	13.73 (14.20)	0.02 (0.03)	—	—	—
Post-K-Pg release and radiate	30.72 (24.70)	0.00 (0.00)	14.47 (13.45)	0.01 (0.01)	—	—	—
Post-K-Pg constraint	9.49 (11.99)	0.06 (0.11)	15.22 (9.99)	0.01 (0.01)	—	—	—
Radiate, post-K-Pg constraint	3.18 (3.60)	0.30 (0.28)	2.57 (2.70)	0.40 (0.40)	—	—	—
OU	9.32 (3.58)	0.01 (0.01)	9.50 (8.39)	0.04 (0.08)	—	—	—
<i>OU shift</i>	28.78 (24.64)	0.00 (0.01)	13.10 (13.06)	0.02 (0.02)	—	—	—
BM	30.94 (45.78)	0.00 (0.01)	15.64 (18.18)	0.08 (0.13)	84.88 (49.60)	0.00 (0.00)	—
BM <i>shift</i>	2.22 (4.22)	0.45 (0.31)	4.51 (8.40)	0.32 (0.25)	10.74 (13.18)	0.32 (0.40)	—
OU	24.78 (47.30)	0.06 (0.14)	17.69 (18.19)	0.03 (0.05)	11.13 (22.50)	0.53 (0.45)	—
Early burst	22.59 (38.93)	0.11 (0.23)	12.63 (14.65)	0.06 (0.07)	22.75 (27.12)	0.04 (0.10)	—
BM with trend	45.74 (17.75)	0.00 (0.00)	85.07 (16.03)	0.00 (0.00)	22.36 (16.84)	0.12 (0.26)	—
White noise	—	—	—	—	—	—	—
<i>Measurement with greatest predictive power</i>							
Post-K-Pg release	16.49 (17.54)	0.01 (0.03)	2.64 (4.11)	0.16 (0.10)	—	—	—
Post-K-Pg release and radiate	18.40 (17.63)	0.01 (0.03)	3.78 (4.02)	0.10 (0.10)	—	—	—
Post-K-Pg constraint	7.07 (11.65)	0.12 (0.15)	5.25 (4.12)	0.06 (0.12)	—	—	—
Radiate, post-K-Pg constraint	1.92 (3.10)	0.41 (0.29)	3.09 (2.35)	0.17 (0.26)	—	—	—
OU	7.07 (3.93)	0.04 (0.03)	4.06 (3.52)	0.12 (0.17)	—	—	—
<i>OU shift</i>	15.91 (17.78)	0.02 (0.05)	2.64 (3.95)	0.16 (0.11)	—	—	—
BM	31.99 (45.42)	0.00 (0.00)	17.61 (16.02)	0.03 (0.07)	83.86 (48.01)	0.00 (0.00)	—
BM <i>shift</i>	4.28 (4.75)	0.23 (0.25)	8.99 (8.13)	0.05 (0.07)	12.06 (13.34)	0.26 (0.37)	—
OU	25.33 (47.04)	0.05 (0.12)	19.96 (16.02)	0.01 (0.03)	9.60 (20.64)	0.55 (0.43)	—
Early burst	22.81 (38.92)	0.11 (0.23)	15.67 (12.88)	0.02 (0.03)	19.71 (25.95)	0.08 (0.17)	—
BM with trend	48.30 (17.43)	0.00 (0.00)	91.68 (16.00)	0.00 (0.00)	23.98 (17.01)	0.12 (0.26)	—
White noise	—	—	—	—	—	—	—

Table S5: AIC and AIC_w scores for model fitting analyses of only extant taxa. Mean values are reported, with standard deviations in parentheses.

Model	ΔAIC	AIC_w
Post-K-Pg release and radiate	4.27 (2.39)	0.06 (0.05)
Post-K-Pg constraint	5.98 (2.40)	0.03 (0.02)
Radiate, post-K-Pg constraint	4.06 (2.49)	0.07 (0.08)
Radiate,constrained	6.26 (2.43)	0.02 (0.02)
OU_{shift}	10.51 (2.95)	0.01 (0.01)
BM_{shift}	4.19 (2.39)	0.06 (0.05)
BM	1.03 (1.62)	0.26 (0.12)
OU	1.73 (1.39)	0.19 (0.14)
Early burst	1.40 (1.91)	0.24 (0.16)
White noise	93.49 (3.81)	0.00 (0.00)

LITERATURE CITED

- Ashmole, N. P., 1968. Body size, prey size, and ecological segregation in five sympatric tropical terns (Aves: Laridae). *Systematic Zoology* 13:292–304.
- Ballman, P., 1983. A new species of fossil barbet (Aves: Piciformes) from the middle Miocene of the Nördlinger Ries (Southern Germany). *Journal of Vertebrate Paleontology* 3:43–48.
- Becker, J. J., 1986. A fossil Motmot (Aves: Momotidae) from the Late Miocene of Florida. *The Condor* 88:478–482.
- Benson, R. B. J., R. A. Frigot, A. Goswani, and R. J. Butler, 2014. Compatition and constraint drove Cope'srule in the evolution of giant flying reptiles. *Nature Communications* 5:doi: 10.1038/ncomms4567.
- Boles, W. E., 1997. A kingfisher (Hacyonidae) from the Miocene of Riversleigh, Northwestern Queensland, with comments on the evolution of kingfishers in Australo-Papua. *Memoirs of the Queensland Museum* 41:229–234.
- Brodkorb, P., 1970. The paleospecies of woodpeckers. *Quarterly Journal of the Florida Academy of Sciences* 33:132–134.
- Butler, R. J., P. M. Barrett, S. Nowbath, and P. Upchurch, 2009. Estimating the effects of sampling biases on pterosaur diversity patterns: implications for hypotheses of bird/pterosaur competitive replacement. *Paleobiology* 35:432–446.
- Clarke, J. A., D. T. Ksepka, A. N. Smith, and M. A. Norell, 2009. Combined phylogenetic analysis of new North American fossil species confirms widespread Eocene distribution for stem rollers Aves, Coracii. *Zoological Journal of the Linnean Society* 157:586–611.
- De Pietri, V. L., A. Manegold, L. Costeur, and G. Mayr, 2011. A new species of woodpecker (Aves; Picidae) from the early Miocene of Saulcet (Allier, France). *Swiss Journal of Palaeontology* 130:307–314.
- DeBee, A. M., 2012. A taxonomic and anatomic assessment of the extinct Zygodactylidae (Aves) from the Green River Formation of Wyoming and placement of Zygodactylidae within Aves. Master's thesis, The University of Texas at Austin.
- Dyke, D. J. and J. H. Cooper, 2000. A new psittaciform bird from the London Clay (Lower Eocene) of England. *Palaeontology* 43:271–285.
- Dyke, G. J. and R. L. Nudds, 2008. The fossil record and limb disparity of enantiornithines, the dominant flying birds of the Cretaceous. *Lethaia* 42:248–254.
- Dyke, G. J. and D. M. Waterhouse, 2001. A mousebird (Aves: Coliiformes) from the Eocene of England. *Journal of Ornithology* 142:7–15.

- Feduccia, A. and D. L. Martin, 1976. The Eocene zygodactyl birds of North America (Aves: Piciformes). *Smithsonian Contributions to Paleobiology* 27:101–110.
- Goodman, S. M., 2010. A description of a new species of Brachypteracias (Family Brachypteraciidae) from the Holocene of Madagascar. *Ostrich* 71:318–322.
- Harrison, C. J. O., 1982. The earliest parrot: A new species from the British Eocene. *Ibis* 124:203–210.
- Houde, P. and L. Olson, 1992. A radiation of coly-like birds from the Eocene of North American; Aves; Sandcoleiformes new order. Pp. 137–160, *in* K. E. Campbell, ed. *Papers in Avian Paleontology Honoring Pierce Brodkorp*. Natural History Museum of Los Angeles County, Science Series 36.
- Ksepka, D. T. and J. A. Clarke, 2009. Affinities of *Palaeospiza bella* and the phylogeny and biogeography of mousebirds. *The Auk* 126:245–259.
- , 2010. New fossil mousebird (Aves: Coliiformes) with feather preservation provides insight into the ecological diversity of an Eocene North American avifauna. *Biological Journal of the Linnean Society* 160:685–706.
- , 2012. A new stem parrot from the Green River Formation and the complex evolution of the grasping foot in Pan-Psittaciformes. *Journal of Vertebrate Palaeontology* 32:395–406.
- Ksepka, D. T., J. A. Clarke, and L. Grande, 2011. Stem parrots (Aves, Halcyornithidae) from the Green River Formation and a combined phylogeny of Pan-Psittaciformes. *Journal of Paleontology* 85:835–852.
- Lack, D., 1946. Competition for food by birds of prey. *Journal of Animal Ecology* 15:123–129.
- Manegold, A. and A. Louchart, 2012. Biogeographic and Paleoenvironmental Implications of a New Woodpecker Species (Aves, Picidae) from the Early Pliocene of South Africa. *Journal of Vertebrate Paleontology* 32:926–938.
- Martin, T. E., 1985. Resource selection by tropical frugivorous birds: integrating multiple interactions. *Oecologia* 66:563–573.
- Mayr, G., 1998a. “Coraciiforme” und “piciforme” Kleinvögel aus dem Mittel-Eozän der Grube Messel (Hessen, Deutschland). *Cour Forsch Inst Senckenberg* 205:1–101.
- , 1998b. A new family of Eocene zygodactyl birds. *Senckenbergiana Lethaea* 78:199–209.
- , 1998c. “Coraciiforme” und “piciforme” Kleinvögel aus dem Mittel-Eozän der Grube Messel (Hessen, Deutschland). *Cour Forsch Inst Senckenberg* 205:1–101.
- , 2000a. A new mousebird (Coliiformes: Coliidae) from the Oligocene of Germany. *Journal of Ornithology* 141:85–92.

- , 2000b. New or previously unrecorded avian taxa from the Middle Eocene of Messel (Hessen, Germany). *Mitt Mus Naturkd Berl Geowiss Reihe* 3:207–219.
- , 2000c. Tiny hoopoe-like birds from the middle Eocene of Messel (Germany). *The Auk* 117:964–970.
- , 2001a. New specimens of the Middle Eocene fossil mousebird *Selmes absurdipes* Peters 1999. *Ibis* 143:427–434.
- , 2001b. The earliest fossil record of a modern-type piciform bird from the late Oligocene of Germany. *Journal of Ornithology* 142:2–6.
- , 2002. A new species of *Plesiocathartes* (Aves: ?Leptosomidae) from the Middle Eocene of Messel, Germany. *PaleoBios* 22:10–20.
- , 2004. New specimens of *Hassiavia laticauda* (Aves: Cypselomorphae) and *Quasisyndactylus longibrachis* (Aves: Alcediniformes) from the Middle Eocene of Messel, Germany. *Cour Forsch Inst Senckenberg* 252:23–28.
- , 2005a. A tiny barbet-like bird from the Lower Oligocene of Germany: the smallest species and earliest substantial fossil record of the Pici (woodpeckers and allies). *The Auk* 122:1055–1063.
- , 2005b. New trogons from the early Tertiary of Germany. *Ibis* 147:512–518.
- , 2005c. The postcranial osteology and phylogenetic position of the Middle Eocene *Messelastur gratulator* Peters, 1994 - a morphological link between owls (Strigiformes) and falconiform birds? *Journal of Vertebrate Paleontology* 25:635–645.
- , 2010. Mousebirds (Coliiformes), parrots (Psittaciformes), and other small birds from the late Oligocene/ early Miocene of the Mainz Basin, Germany. *Neues Jahrbuch für Geologie und Paläontologie, Abhandlungen* 258:129–144.
- , 2013. Late Oligocene mousebird converges on parrots in skull morphology. *Ibis* 155:384–396.
- Mayr, G. and M. Daniels, 1998. Eocene parrots from Messel (Hessen, Germany) and the London Clay of Walton-on-the-Naze (Essex, England). *Senckenbergiana Lethaea* 78:157–177.
- Mayr, G. and U. B. Göhlich, 2004. A new parrot from the Miocene of Germany, with comments on the variation of hypotarsus morphology in some Psittaciformes. *Belgian Journal of Zoology* 134:47–54.
- Mayr, G. and R. Gregorová, 2012. A tiny stem representative of Pici (Aves, Piciformes) from the early Oligocene of the Czech Republic. *Paläontologische Zeitschrift* 86:333–343.
- Mayr, G. and C. W. Knopf, 2007. A tody (Alcediniformes: Todidae) from the early Oligocene of Germany. *The Auk* 124:1294–130.

- Mayr, G. and N. Micklich, 2010. New specimens of the avian taxa *Eurotrochilus* (Trochilidae) and *Palaeotodus* (Todidae) from the early Oligocene of Germany. *Paläontologische Zeitschrift* 84:387–395.
- Mayr, G. and C. Mourer-Chauviré, 2000. Rollers (Aves: Coraciiformes s.s.) from the Middle Eocene of Messel (Germany) and the Upper Eocene of the Quercy (France). *Journal of Vertebrate Paleontology* 20:533–546.
- Mayr, G., C. Mourer-Chauviré, and I. Weidig, 2004. Osteology and systematic position of the Eocene Primobucconidae (Aves, Coraciiformes *sensu stricto*), with first records from Europe. *Journal of Systematic Palaeontology* 2:1–12.
- Mayr, G. and D. S. Peters, 1998. The mousebirds (Aves: Coliiformes) from the Middle Eocene of Grube Messel (Hessen, Germany). *Senckenberg Lethaea* 78:179–197.
- Mayr, G., R. S. Rana, K. D. Rose, A. Sahni, K. Kumar, L. Singh, and T. Smith, 2010. *Quercypsitta*-like birds from the Early Eocene of India (Aves, ?Psittaciformes). *Journal of Vertebrate Palaeontology* 30:467–478.
- Mayr, G. and T. Smith, 2013. Galliformes, Upupiformes, Trogoniformes, and other avian remains (?Phaethontiformes and ?Threskiornithidae) from the Rupelian stratotype in Belgium, with comments on the identity of “*Anas*” *benedeni* Sharpe, 1899. *Paleornithological Research* 306:23–35.
- Mayr, G. and N. Zelenkov, 2009. New specimens of zygodactylid birds from the middle Eocene of Messel, with description of a new species of *Primozygodactylus*. *Acta Palaeontologica Polonica* 54:15–20.
- Mlíkovský, J., 1998. A new parrot (Aves: Psittacidae) from the Early Miocene of the Czech Republic. *Acta Societatis Zoologicae Bohemicae* 62:335–341.
- Mlíkovský, J. and U. B. Göhlich, 2000. A new wood-hoopoe from the Early Miocene of Germany and France. *Acta Societatis Zoologicae Bohemicae* 64:419–424.
- Morlo, M., S. Schaal, G. Mayr, and C. Seiffert, 2004. An annotated taxonomic list of the Middle Eocene (MP 11) Vertebrata of Messel. *Courier Forschungsinstitut Senckenberg* 252:95–108.
- Mourer-Chauviré, C., 1992. Une nouvelle famille de perroquets (Aves, Psittaciformes) dans l’Éocène supérieur des Phosphorites du Quercy, France. *Geobios Mem Spec* 14:169–177.
- Mourer-Chauviré, C., J.-B. Peyrouse, and M. Hugueney, 2013. A new roller (Aves: Coraciiformes s.s.:Coraciidae) from the Early Miocene of Saint-Gérang-le-Puy area, Allier, France. *Palaeornithological Research* Pp. 81–92.
- Olson, S. L., 1975. Paleornithology of St Helena Island, south Atlantic Ocean. *Smithsonian Contributions to Paleobiology* 23:32–35.
- Santa Rosalia reconsidered: size ratios and competition, 1981. *Evolution* 35:1206–1228.

- Schoener, T. W., 1974. Resource partitioning in ecological communities. *Science* 185:27–39.
- Steadman, D. W., 2006. A new species of extinct parrot (Psittacidae: *Electus*) from Tonga and Vanuatu, South Pacific. *Pacific Science* 60:137–145.
- Waterhouse, D. M., B. E. K. Lindow, N. V. Zelenkov, and G. J. Dyke, 2008. Two new parrots (Psittaciformes) from the Lower Eocene Fur Formation of Denmark. *Palaeontology* 51:575–582.
- Weidig, I., 2006. The first New World occurrence of the Eocene bird *plesiocathartes* (aves: ?leptosomidae). *Paläontologische Zeitschrift* 80:230–237.
- , 2010. New birds from the Lower Eocene Green River formation North America. *Records of the Australian Museum* 62:29–44.
- Werner, E. E. and J. F. Gilliam, 1984. The ontogenetic niche and species interactions in size-structured populations. *Annual Review of Ecology and Systematics* 15:393–425.
- Wilson, D. S., 1975. The adequacy of body size as niche difference. *The American Naturalist* 109:769–784.
- Worthy, T. H., A. J. D. Tennyson, and P. R. Scofield, 2011. An Early Miocene diversity of parrots (Aves, Strigopidae, Nestorinae) from New Zealand. *Journal of Vertebrate Paleontology* 31:1102–1116.