Supporting Information

Single-cell visualization indicates direct role of sponge host in uptake of dissolved organic matter

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Figure S1

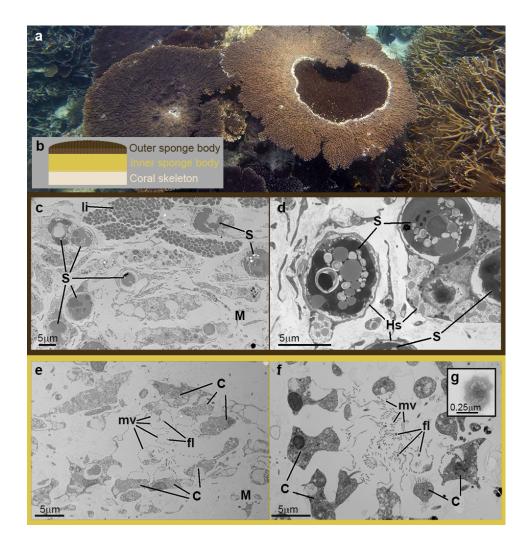


Figure S1: Macro and microscopic observations of the bioeroding sponge *Cliona orientalis* from the Great Barrier Reef, Australia. **a** *C. orientalis* (brown patch) in competition with a plating Acroporid coral. **b** Vertical cross section of the experimental sponge cylinders (35 mm diameter, colour scheme applies to figures of the main article). The outer sponge body hosts most of the endosymbiotic dinoflagellates of the genus *Gerakladium*, but the inner sponge body where light does not penetrate is relatively void of *Gerakladium*. The underlying coral skeleton has not yet been eroded by the sponge. **c** Transmission Electron Micrograph (TEM) of the outer sponge body, showing the dinoflagellate symbiont cells (S) and dense lipid accumulations (li) in the sponge mesohyl (M). **d** Further magnification shows the intracellular positioning of dinoflagellate symbionts (S) inside archaeocyte-like host cells (Hs). **e** TEM image of the inner sponge body showing the heterotrophic feeding cells or choanocytes (C) that line the aquiferous canals. Choanocytes bear a flagellum (fl, seen in cross section), which is surrounded by a collar of cellular extensions called microvilli (mv). Beating of the flagellae creates a water flow across the microvilli enabling trapping of food particles which are later phagocytosed by the cell, or passed on to other cells in the surrounding mesohyl (M). **f** Further magnification showing a cross section of choanocytes forming a so-called choanocyte chamber. **g** TEM image of an unidentified bacterium (potentially *Rhodothalassium* sp., an Alphaproteobacterium; Ramsby et al, 2018, *Mol Ecol* 27:2124–2137) found occasionally in the inner but also the outer sponge body.

Figure S2

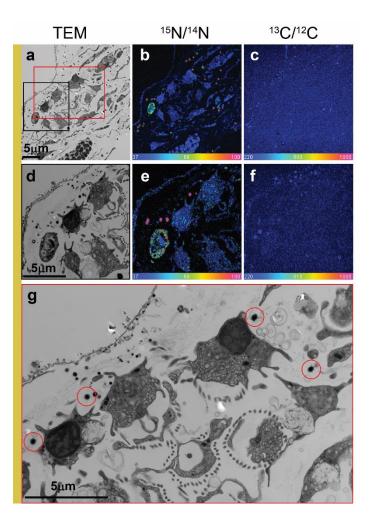


Figure S2: Visualization of ¹⁵N and ¹³C isotopic distribution in the inner body of the *Cliona orientalis* holobiont at the end of the chase period (t=24h). **a, d, g** TEM displaying sponge choanocytes and other host material, as well as unidentified prokaryotic microorganisms (circled in red in **g**, also shown in higher resolution in Fig. S1g). Selected areas displayed in **a** are enlarged in **d** (black rectangle) and in **g** (red rectangle). **b**, **e** Corresponding ¹⁵N-distribution showing enriched prokaryotic microorganisms ($\Delta\delta^{15}N = 1684 \pm 107\%_0$, mean \pm SEM of the 27 microorganisms detected in **b**), and other sponge material. **c,f** Corresponding ¹³C-distribution of the same microorganisms ($\Delta\delta^{13}C = 120 \pm 22\%_0$). Note that in this case the carbon enrichment is substantially less profound than the nitrogen enrichment. Images **b**, **c**, **e**, and **f** show the deviation of the ratios from the natural abundance ratio, using a rainbow scale in hue saturation intensity. The rainbow scale ranges from natural abundance in blue (0.0037 for ¹⁵N/¹⁴N and 2x 0.0110 for ¹³C/¹²C) to several-fold enrichment above natural abundance in red (approx. 3-fold for ¹⁵N/¹⁴N and 5-fold for ¹³C/¹²C). The ochre yellow bar on the far left of the panel refers to the colour scheme in Fig. S1b.

Table S1

Results of two-factorial permutational analysis of variance testing the Euclidean dissimilarities between the ${}^{15}N/{}^{14}N$ and ${}^{13}C/{}^{12}C$ ratios of each area of interest [AOI, 1st factor with 5 levels: (I) Dinoflagellate symbionts=S, (II) Cytoplasm of dinoflagellate-hosting sponge cells=Hs, (III) nearby outerbody mesohyl cells or intercellular areas=Mo, (IV) Choanocytes=C and (V) nearby inner-body mesohyl cells and intercellular space=Mi] between the three time-points (Time, 2nd factor with 3 levels: 0h, 3h and 24h). Bold face indicates statistical significance (P < 0.05). Dispersions differed amongst the compared groups, with the greatest dispersion found in the largest group, rendering the permutational tests relatively conservative (Anderson and Walsh, 2013, *Ecol Monograph* 83: 557-574). df=degrees of freedom; SS=sum of squares; MS=mean sum of squares; Pseudo-F=F value by permutation, P_(perm)=P-value by permutation.

	Main Permanova Test						Pairwise Comparisons			
Parameter	Source	df	SS	MS	Pseudo-F	P _(perm)	Level	Groups	Pseudo-T	P _(perm)
¹⁵ N/ ¹⁴ N	Time	2	3.14E-03	1.57E-03	3.4404	0.0380	S	0h versus 3h	4.6753	0.0002
	AOI	4	7.85E-03	1.96E-03	4.3003	0.0108		0h versus 24h	4.1256	0.0006
	Time x AOI	8	1.54E-02	1.92E-03	4.2036	0.0036		3h versus 24h	2.4791	0.0146
	Res	722	0.32969	4.57E-04			Hs	0h versus 3h	2.2993	0.0004
								0h versus 24h	3.9715	0.0010
								3h versus 24h	2.8916	0.0042
							Мо	0h versus 3h	9.1998	0.0002
								0h versus 24h	3.6583	0.0002
								3h versus 24h	2.6775	0.0054
							С	0h versus 3h	4.0446	0.0002
								0h versus 24h	8.0880	0.0002
								3h versus 24h	2.8360	0.0042
							Mi	0h versus 3h	8.2188	0.0002
								0h versus 24h	5.1020	0.0002
								3h versus 24h	0.5716	0.5686
¹³ C/ ¹² C	Time	2	1.11E-04	5.55E-05	2.6311	0.0738	S	0h versus 3h	2.4770	0.0154
	AOI	4	5.13E-04	1.28E-04	6.0761	0.0154		0h versus 24h	2.4800	0.0176
	Time x AOI	8	6.20E-04	7.75E-05	3.6694	0.0246		3h versus 24h	0.8709	0.3790
	Res	722	1.52E-02	2.11E-05			Hs	0h versus 3h	2.0907	0.0420
								0h versus 24h	2.7301	0.0096
								3h versus 24h	0.6242	0.5370
										4

Мо	0h versus 3h	3.4711	0.0012
	0h versus 24h	2.0989	0.0384
	3h versus 24h	2.0808	0.0396
С	0h versus 3h	4.4125	0.0002
	0h versus 24h	8.6999	0.0002
	3h versus 24h	2.7601	0.0044
Mi	0h versus 3h	2.2346	0.0276
	0h versus 24h	4.3404	0.0002
	3h versus 24h	1.6421	0.1088

Table S2

Results of permutational analysis of variance testing the Euclidean dissimilarities between the levels of ¹⁵N- and ¹³C-enrichment ($\Delta\delta$ values above background) of each area of interest (AOI abbreviations as explained in the legend of Table S1) between treatment time-points (3h and 24h). Bold face indicates statistical significance (P < 0.05). Dispersions differed amongst the compared groups, with the greatest dispersion found in the largest group, rendering the permutational tests relatively conservative (Anderson and Walsh, 2013, *Ecol Monograph* 83: 557-574). df=degrees of freedom; SS=sum of squares; MS=mean sum of squares; Pseudo-F=F value by permutation, P_(perm)=P-value by permutation.

Main Permanova Test						Pairwise Comparisons (3h versus 24h)			
Parameter	Source	df	SS	MS	Pseudo-F	P(perm)	AOI Level	Pseudo-T	P(perm)
¹⁵ N-enrichment	Time	1	9.97E+06	9.97E+06	0.2169	0.6412	S	2.4791	0.0122
	AOI	4	1.07E+09	2.68E+08	5.8416	0.0006	Hs	2.8916	0.0042
	Time x AOI	4	7.74E+08	1.93E+08	4.2106	0.0048	Мо	2.6775	0.0044
	Res	534	2.45E+10	4.60E+07			С	2.8360	0.0056
							Mi	0.57164	0.5738
¹³ C-enrichment	Time	1	1.23E+05	1.23E+05	1.9340	0.1630	S	0.8709	0.3842
	AOI	4	1.63E+06	4.07E+05	6.3833	0.0004	Hs	0.6242	0.5298
	Time x AOI	4	7.25E+05	1.81E+05	2.8463	0.0322	Мо	2.0808	0.0380
	Res	534	3.40E+07	63698			С	2.7601	0.0026
							Mi	1.6421	0.0980