## Supplementary Information

## Supplementary Methods

## Whole-mount immunostaining and in situ hybridization in Chondrichthyans

For Pax3 and Lbx1 expression, 12 replicates for each gene were investigated. For Wnt3, Cyp26a1, Fgf8, and Hox genes, 3 embryos for each gene were investigated. The nerve staining method was modified from a previous study (50). Briefly, skate and shark embryos were transferred from salt water to $500 \mathrm{mg} / \mathrm{L}$ Tricaine-S and then fixed in $4 \%$ PFA overnight. The following day, embryos were washed in phosphate buffered saline (PBS) containing 1\% Triton (PBTriton) for 3 hours. Embryos were incubated in $0.25 \%$ trypsin/PBS for 5 minutes and immersed in pre-cooled acetone at $-20^{\circ} \mathrm{C}$ for 10 minutes. After brief rinsing with PBTriton, specimens were placed in blocking solution (PBTriton containing $10 \%$ goat serum, $1 \%$ dimethyl sulfoxide, and $5 \% \mathrm{H} 2 \mathrm{O} 2$ ) at $4^{\circ} \mathrm{C}$ overnight. The following day, the solution was replaced with blocking solution containing 1:50 3A10 antibody for nerve staining (Developmental Studies Hybridoma Bank) or 1:70 myosin heavy chain for muscle staining (Developmental Studies Hybridoma Bank). Embryos were incubated in antibody solution at $4^{\circ} \mathrm{C}$ for 72 hours. Embryos were washed with PBTriton for 5 hours and then incubated with blocking solution containing 1:1000 peroxidase-conjugated secondary antibody (Jackson Laboratory, catalogue number 115-035-003). The next day, embryos were washed with PBTriton for 5 hours and subjected to DAB color development. At each stage, we stained nerves of 6 embryos and muscles of 4 embryos.

## Supplementary Figure 1 | The dual contribution of Pax3-positive muscle precursor cells into the pectoral fin and body wall muscles.

(A) Pax3-positive cells migrated into the pectoral fin (arrowheads) and body wall muscles (arrows) in skate embryos at stage 29. Scale bar is 1 mm .

## Supplementary Figure 2 | The development of hypaxial muscles in shark embryos at stage 30 .

(A) Ventral view of shark muscles stained by myosin heavy chain antibody at stage 30 . Note that pectoral fin muscles (arrow) and body wall muscles (arrowheads) develop at the same axial level. Scale bar is 1 mm .

Supplementary Figure 3 | The pectoral and pelvic fin domains in wildtype and RA-treated embryos.
(A) The pectoral fin (dark blue) develops in the lateral plate mesoderm adjacent to somites 1-36 (or 37) and pelvic fins (light blue) from 36 (or 37 ) to 50 . We counted observable somites under the microscope. In RA-treated embryos, the pectoral fins were narrower than wildtype (somites 1-28). The severity of phenotypes varies, however, all embryos showed the same phenotype.

## Supplementary Figure 4 | Hoxa9 expression patterns of embryos treated by retinoic acid from stage 23 to 24.

Whole-mount in situ hybridization of $L$. erinacea embryos at stage 23 to 24. (A) Hoxa9 expression in a wild type embryo. LPM expression starts from somite 26 (arrowhead).
(B) Hoxa9 expression in an embryo treated by RA. LPM expression starts from somite 23 (arrowhead). Note that Hoxa9 expression is shifted anteriorly in the RA+ embryo by 3 somites. Scale bars are 1 mm .

## Supplementary Figure 5 |. Neurofilament staining of RA-treated embryos

(A) The brachial plexus did not develop in embryos treated by RA, although nerves innervate hypobranchial and pectoral muscles (arrows). A scale bar is 1 mm .

Supplementary Table 1 | Cloning primers for Cdx2, Cyp26a1, Lbx1, Pax3, and Hox genes of skate



A


## Supplementary Figure 3



Supplementary Figure 4


Supplementary Figure 5

| Gene | Forward Sequence | Reverse Sequence | Length |
| :--- | :--- | :--- | :--- |
| Hoxa2 | CAAACCATCCCTAGCCTGAA | TGGACCCATGTATTGCTGAA | 1133 bp |
| Hoxa3 | CTCAATCCAGCACAAGCAAA | GGCTGGAGTCCATGAAAAGA | 1104 bp |
| Hoxa4 | CCCCAATTACACTGGAGGAG | GGCAGTTTGTGATCTTTCTTCC | 209 bp |
| Hoxa5 | TAGACGCACAAACGACCAAG | CAGGCAACAGTACGACCAGA | 1053 bp |
| Hoxa9 | GCTGCCCTTACACCAAACAT | TTTCTTTGCACCACAACACC | 975 bp |
| Hoxa10 | ACCTTATTGCCGTTCTGGTG | TTTTCTCCCCAATTCACTCG | 803 bp |
| Hoxa11 | GTTTAGCTGTGCGAATGCAA | TGCGCTTCTTATCGCCTTAT | 1391 bp |
| Hoxa13 | GAACTTCACCGCAAACCAAT | CGTGCAGGCAGAAATGTAAA | 917 bp |
| Hoxd1 | GTCACCAGGGCTGTGACTTT | AGCGTTCGCTATTTCCACAC | 469 bp |
| Hoxd3 | TCAGACATCCAGTGCCAGAG | TCCCTGAGCTGTGTGATGAG | 1002 bp |
| Hoxd4 | ATCCCAAATTTCCTCCTTGC | GGGTTCCCCTCCAGTGTAAT | 400 bp |
| Hoxd8 | AAGCCAGGAGAAGCCCTAAG | TTCTATCCGGCGGTTGATAC | 929 bp |
| Cdx2 | TTTGTCAGCTTTCGCTCCTT | ATGTACGTGAGYTAYCYT | 623 bp |
| Cyp26a1 | CAATGCTCGACATGGTCTTG | CTTCACAGTGGAGCTGGTCA | 552 bp |
| Lbx1 | CCGTTCAGCATCGAGGATA | GCCTTCATCTCTTCCAGGTC | 473 bp |
| Pax3 | ATGAAGGGTGGGGTGGATA | ATCTGGGACTGAACGTGTCC | 1505 bp |
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## Supplementary Table 1

